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(54) **VACCINE**

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CPC **C07K 14/33** (2013.01); **A61K 39/08**
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CPC ... **A61K 38/00**; **A61K 38/4893**; **C07K 14/47**
See application file for complete search history.

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(57) **ABSTRACT**

There is provided a NetB epitope polypeptide comprising at least 10 contiguous amino acids from SEQ ID NO:1 and comprising a mutation in at least one position between amino acids 130 and 297 as compared with the equivalent position in SEQ ID NO:3, the mutation preferably being located within a rim domain, the polypeptide being capable of binding an antibody which binds to SEQ ID NO:1 and having reduced toxicity compared with the toxicity of SEQ ID NO:1. The polypeptide is useful to vaccinate a subject against infection by *Clostridium perfringens*.

8 Claims, 11 Drawing Sheets

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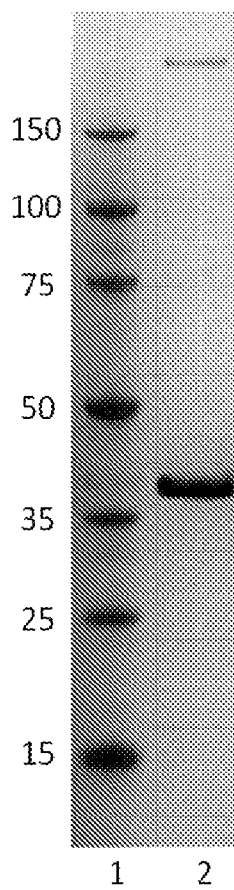


Figure 1

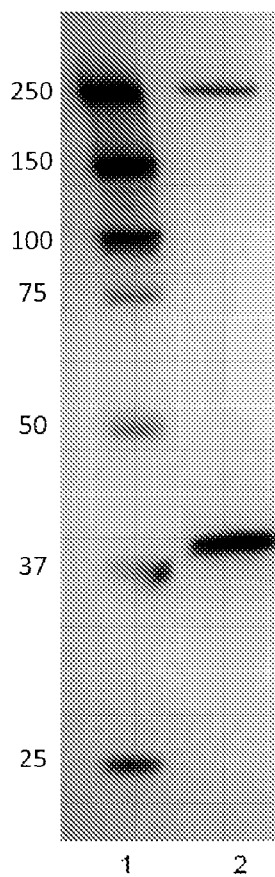


Figure 2

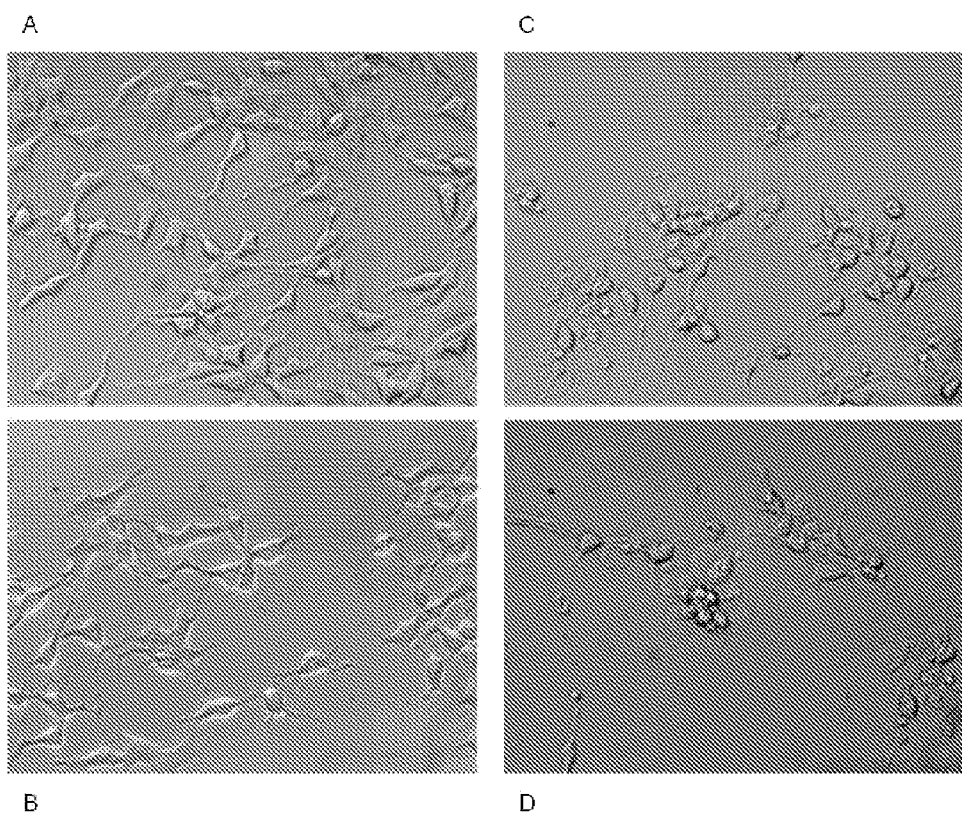


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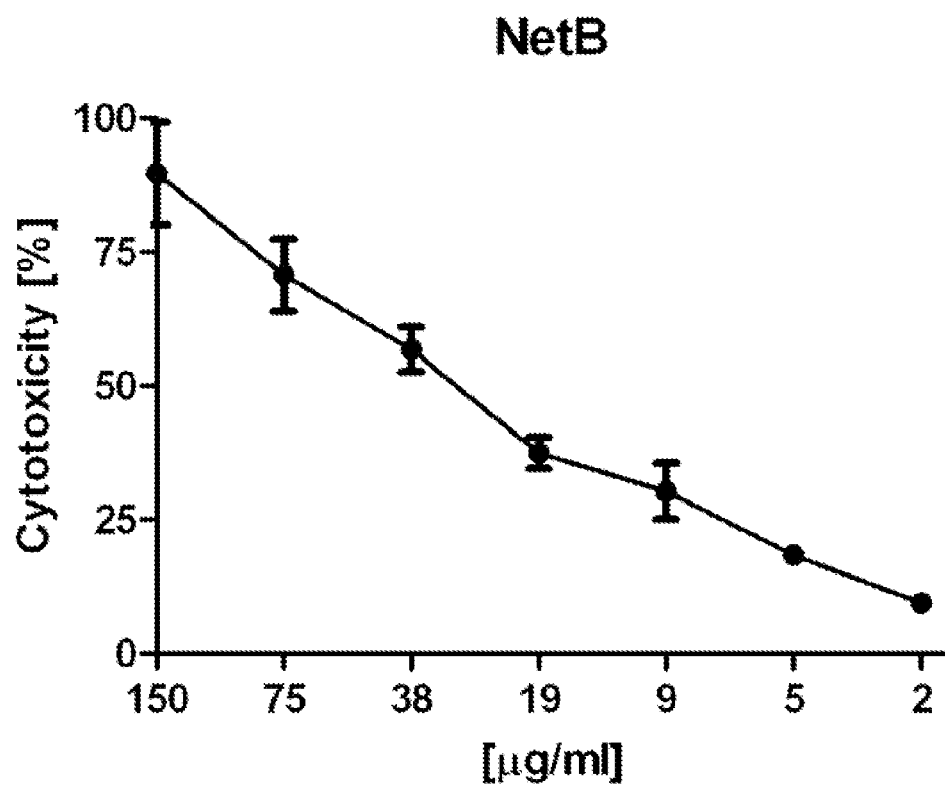


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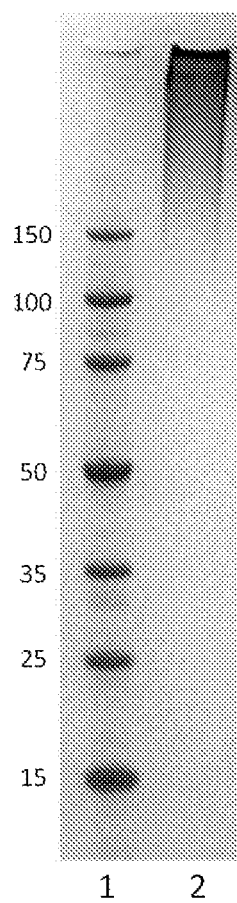
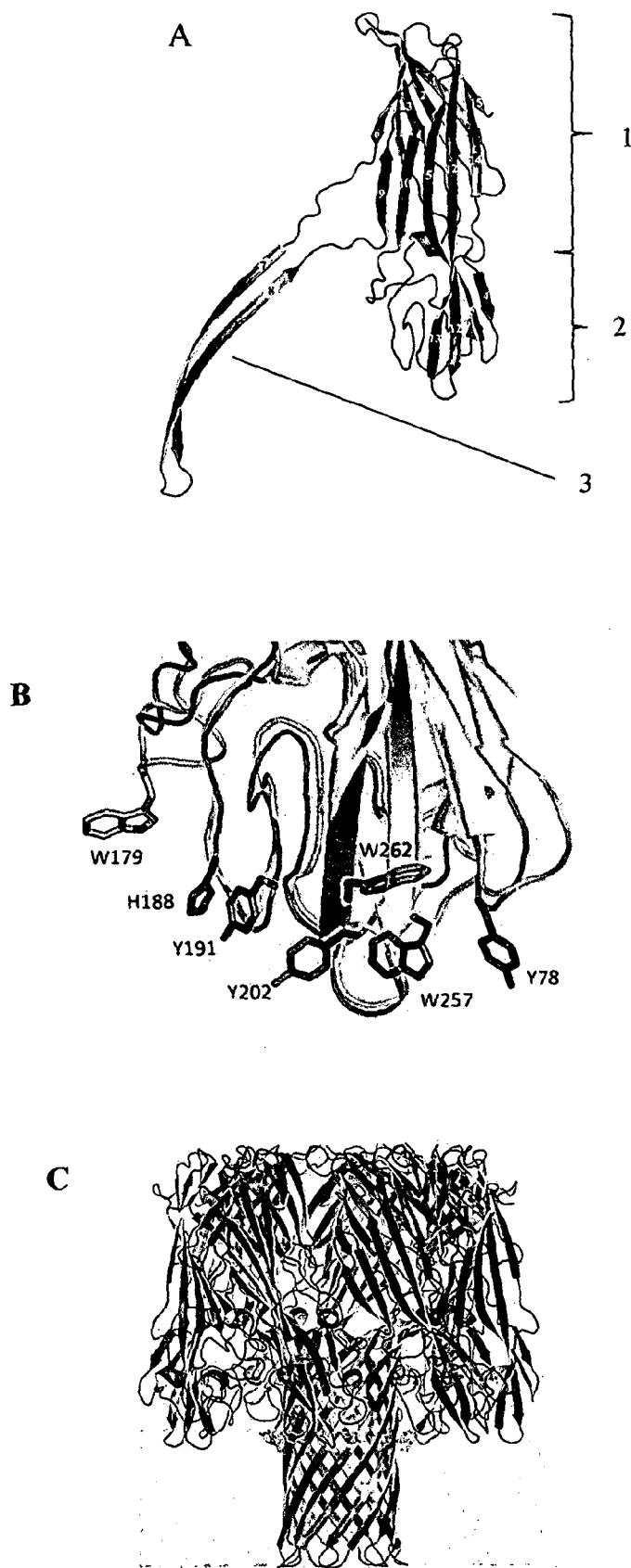


Figure 5

Figure 6



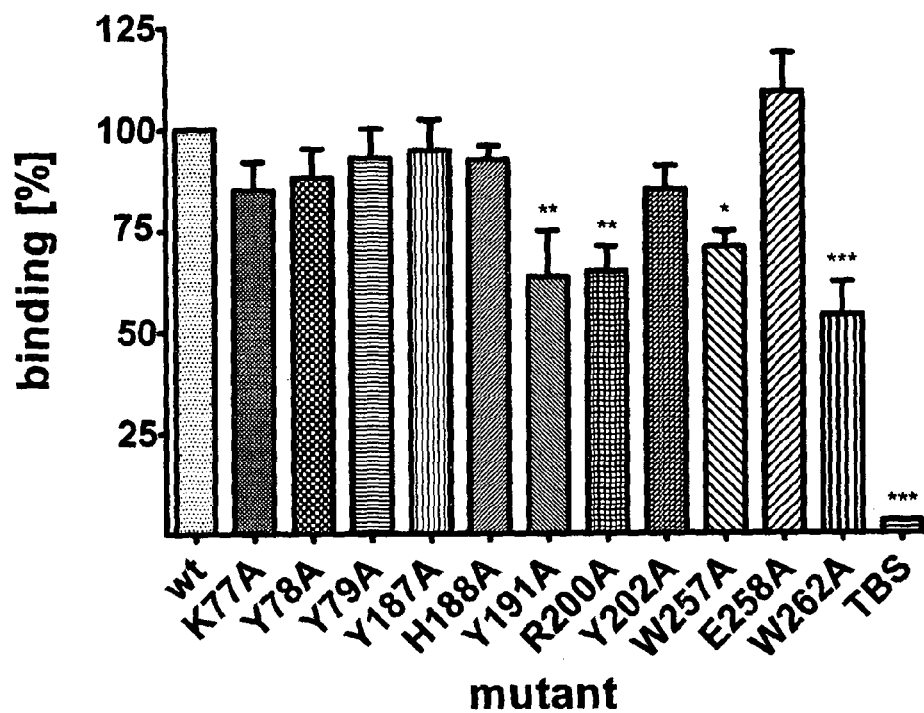


Figure 7A

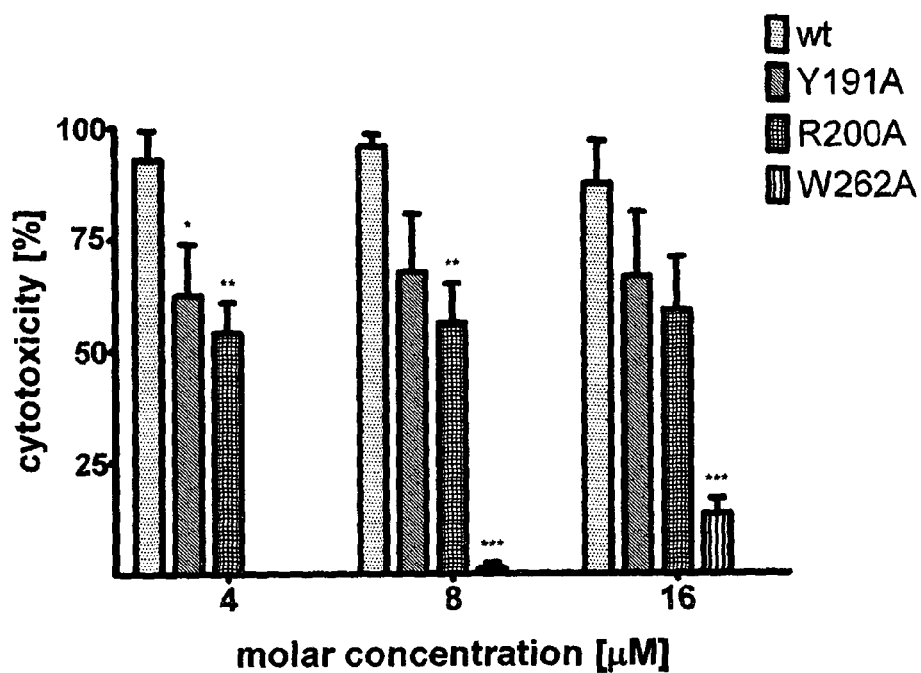


Figure 7B

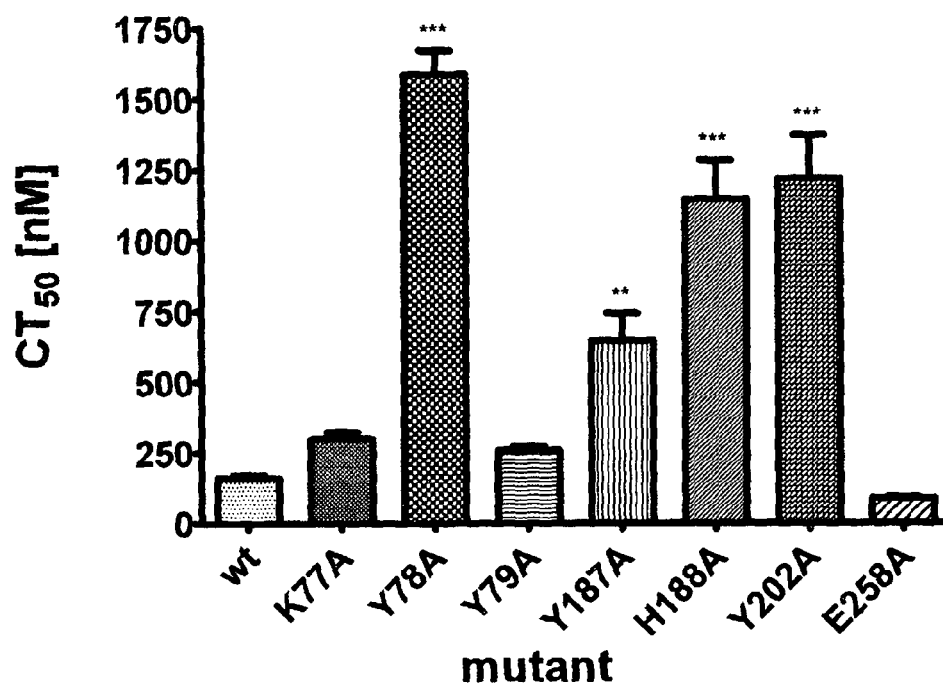


Figure 7C

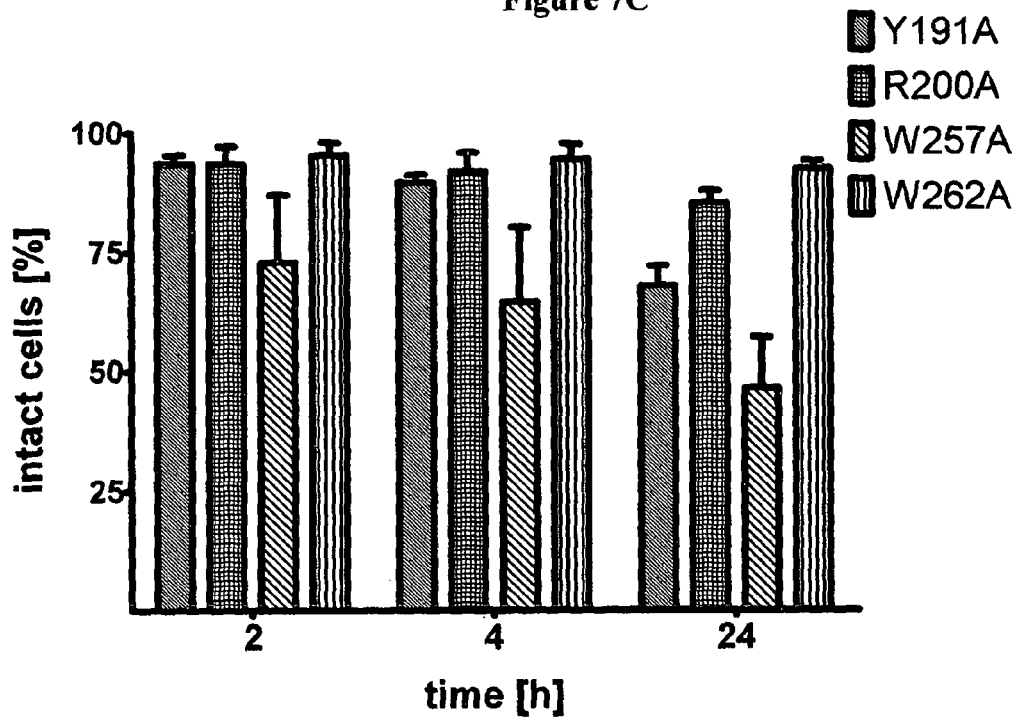


Figure 7D

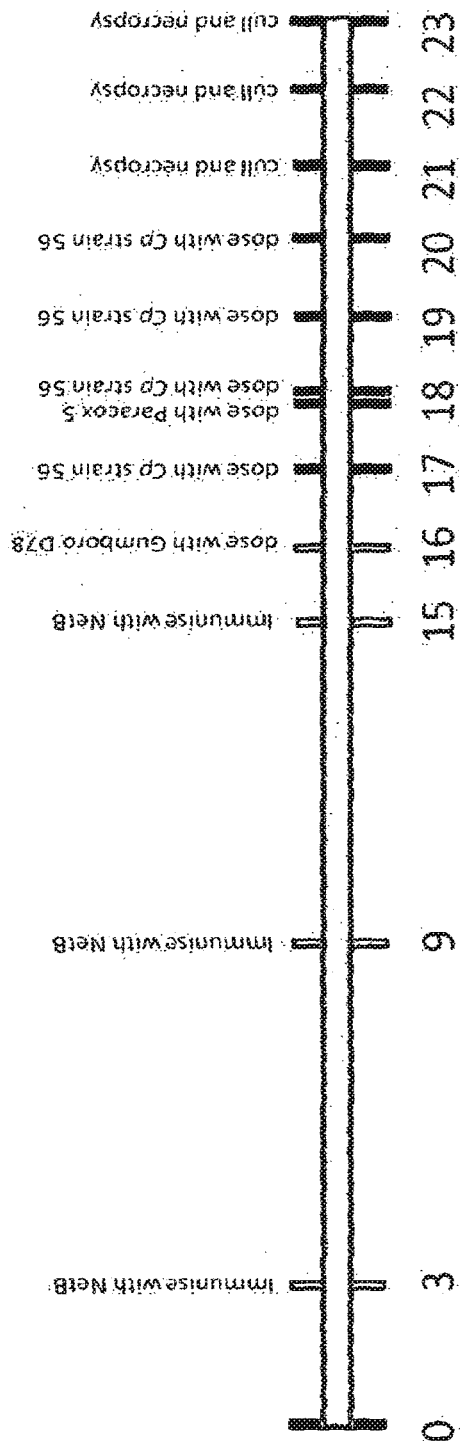
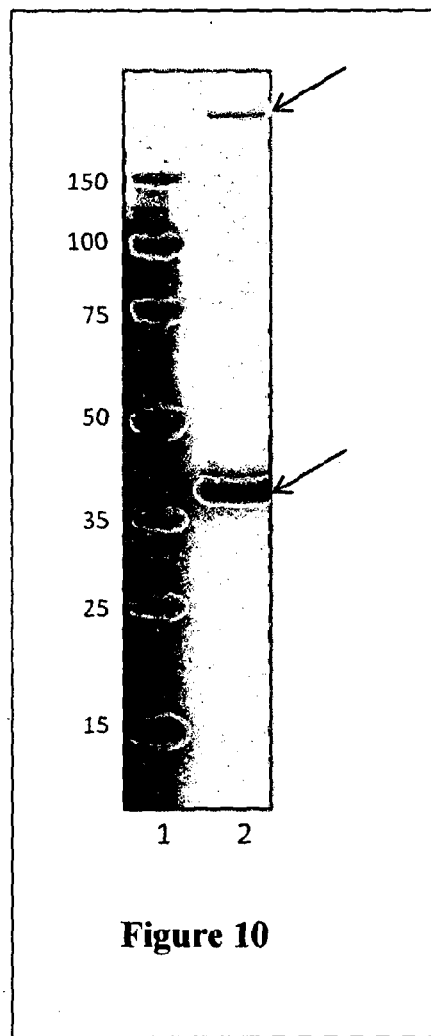
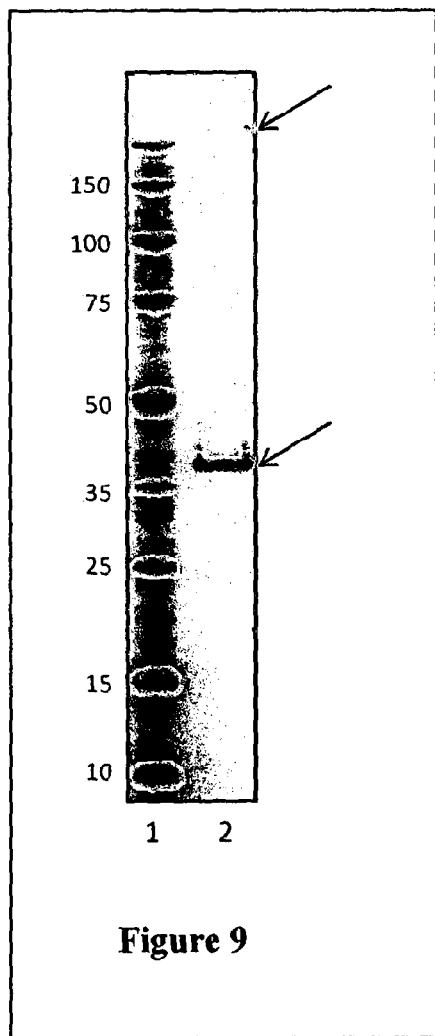


Figure 8



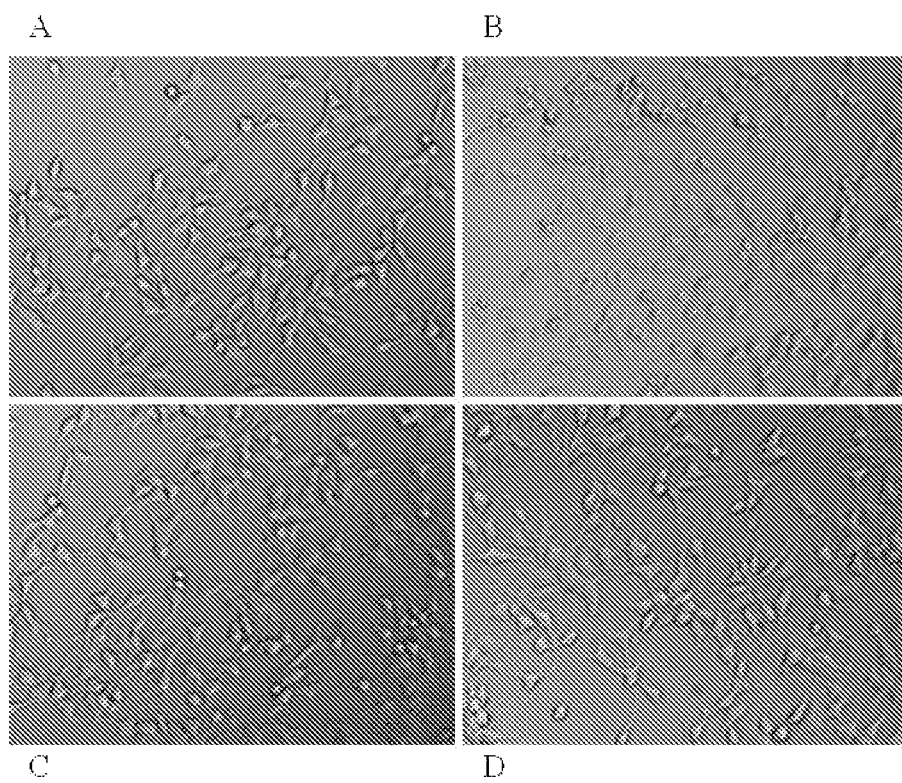


Figure 11

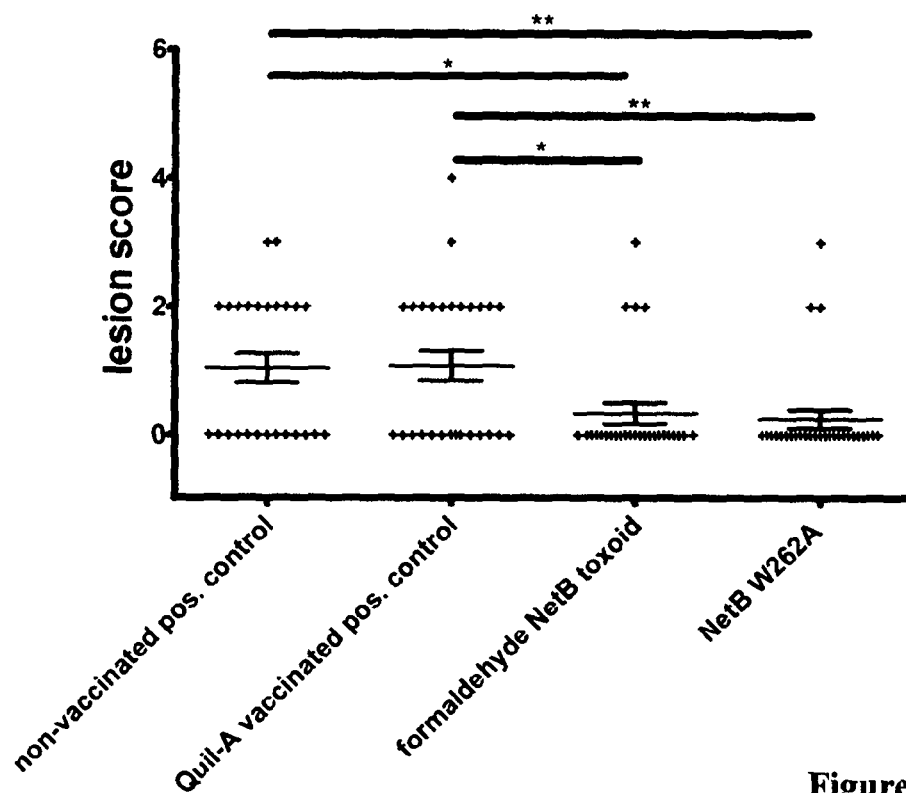
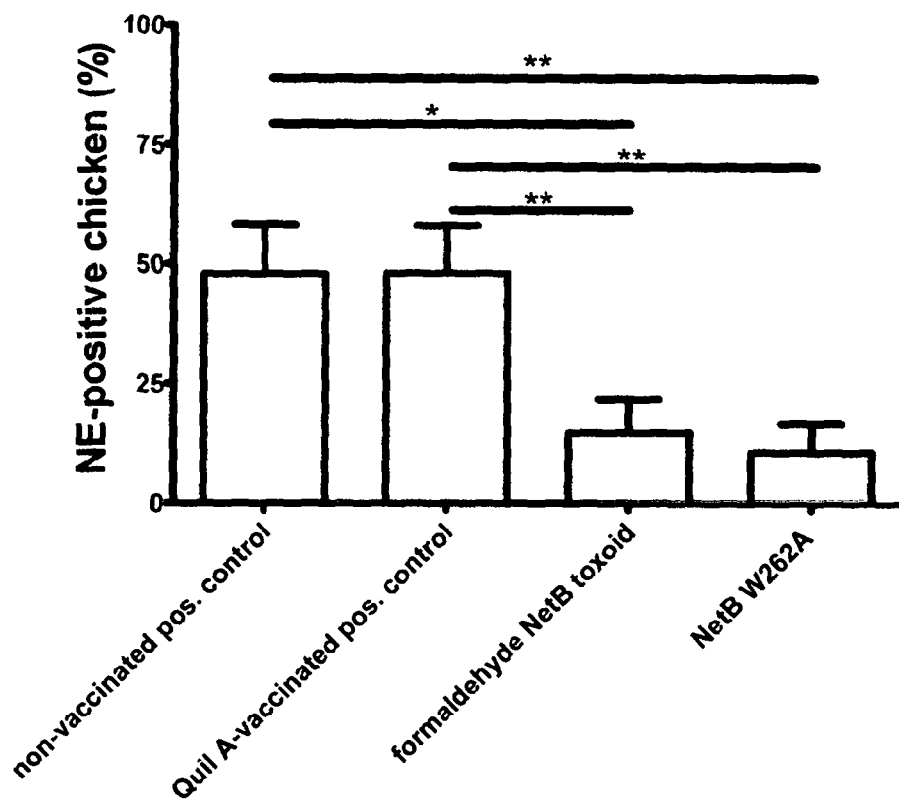


Figure 12



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VACCINE

FIELD OF THE INVENTION

The invention relates to novel polypeptides useful as a vaccine against *Clostridium perfringens*, particularly in chickens and other poultry.

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is the national stage application of International patent application No. PCT/GB2012/052639, entitled "Vaccine," and filed on Oct. 24, 2012, which claims priority to GB patent application No. 1118394.4, entitled "Vaccine" and filed on Oct. 25, 2011, which are hereby incorporated by reference herein in their entireties.

BACKGROUND

Clostridium perfringens is an ubiquitous bacterium that can colonise a variety of different biotopes. Due to its anaerobic lifestyle it is not surprising to find *C. perfringens* as a commensal of the normal gut flora in humans and domesticated animals. However, under certain circumstances it is known to be responsible for causing some severe diseases due to its production of a wide range of toxins (Songer (1996) *Clin Microbiol Rev* vol 9: 216-234). Apart from the four toxins used for typing *C. perfringens* (alpha-, beta-, epsilon-, iota-toxin) it is able to produce a selection of non-typing toxins, such as enterotoxin or perfringolysin O (Petit et al., (1999) *Trends Microbiol* vol 7: 104-110). Recently, a novel toxin named NetB (Necrotic enteritis toxin B) has been identified and suggested to play a role in the pathogenesis of avian necrotic enteritis (NE), a severe gastro-intestinal disease that manifests in gross lesions within the intestines of poultry (WO2008/148166). NE is a re-emerging disease that is causing enormous economic costs to the worldwide poultry industry (around 2 billion US dollars per year) (Keyburn et al., (2008) *PLoS Pathog* vol 4: e26). Its re-emergence is due to the initiative of some governments to prohibit the use of antimicrobial growth promoters in animal feed, amongst others to reduce the evolving spread of antibiotic-resistant bacteria in the environment.

The NetB is produced by *C. perfringens* toxinotype A strains and, to a lesser extent, by strains of type C (Kaldhusdal et al. (1999) *FEMS Immunol Med Microbiol* vol 24: 337-343). The protein is 322 amino acids long in its active form and has an estimated molecular weight of 36.5 kDa. Although the molecular basis of toxicity is still little understood, several studies suggest that the NetB is a new member of the small β -pore-forming toxins (β -PFTs) as it is able to form pores on membranes and shares amino acid sequence similarity with several other related members of the small pore-forming toxins family (38% identity with the beta toxin from *C. perfringens*, 40% identity with the *C. perfringens* delta toxin, and 31% identity with the alpha toxin from *S. aureus*) (Keyburn et al. (2008) *PLoS Pathog* vol 4: e26; Manich et al. (2008) *PLoS One* vol 3: e3764). It was initially assumed that the alpha toxin, which is produced by the same bacterium, is the major virulence factor for causing NE, but experiments with an alpha toxin mutant showed that this strain was still virulent and able to cause disease (Keyburn et al. (2006) *Infect Immun* vol 74: 6496-6500). In contrast, a netB mutant was not capable of causing NE, whereas the wild type and the complemented mutant could (Keyburn et

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al. (2008) *PLoS Pathog* vol 4: e26). However, it is still unsettled as to whether the NetB is the key virulence factor for causing NE, as in some cases it was reported that even *C. perfringens* strains without the netB gene were still capable of virulence (Cooper & Songer (2009) *Vet Microbiol* vol 142: 323-328). Moreover, immunization studies with alpha toxin and other antigens, such as a hypothetical zinc metalloprotease and a pyruvate-ferredoxine oxidoreductase, have been identified to moderately protect chicken from developing NE (Cooper et al. (2009) *Vet Microbiol* vol 133: 92-97; Zekarias et al. (2008) *Clin Vaccine Immunol* vol 15: 805-816; Kulkarni et al. (2010) *Clin Vaccine Immunol* vol 17: 205-214; Kulkarni et al. (2007) *Clin Vaccine Immunol* vol 14: 1070-1077).

The heptameric structure of one of the most widely studied β -PFT, *S. aureus* α -hemolysin (α HL), was determined over 20 years ago (Song L et al. (1996) *Science* vol. 274: 1859-1866) and was, until recently, the only high resolution structure of a β -PFT in the membrane-inserted form. The ring-shaped complex resembles a mushroom with the cap forming the extracellular domain and the stem forming the membrane-spanning region, in which each subunit contributes one β -hairpin. Although NetB appears to form pores in target cell membranes, little is known about the molecular basis for this toxicity which hinders the development of effective control measures against NE.

Several attempts have been made on the development of an effective vaccine to protect chicken against NE but, to date, without significant success.

SUMMARY OF INVENTION

According to a first aspect of the invention, there is provided a NetB epitope polypeptide comprising at least 10 contiguous amino acids from SEQ ID NO:1 and comprising a mutation in at least one position between amino acids 130 to 297 as compared with the equivalent position in SEQ ID NO:3 (or SEQ ID NO:1), the polypeptide being capable of binding an antibody which binds to the polypeptide of sequence SEQ ID NO:1 and having reduced toxicity compared with the toxicity of the polypeptide of sequence SEQ ID NO:1.

This skilled person is readily able to determine "equivalent positions" between two sequences, by aligning sequences to achieve maximum identical amino acids at as many positions as possible, for example by using a global sequence alignment program such as is available via <http://blast.ncbi.nlm.nih.gov/Blast.cgi>, discussed further below.

The inventors have made several polypeptides derived from NetB and having at least one mutation in an amino acid position equivalent to the position in wild-type NetB sequence SEQ ID NO:3 (or SEQ ID NO:1) which, surprisingly, have reduced or absent toxicity compared to the toxicity of the mature protein SEQ ID NO:1, which lacks the N-terminal 30 amino acid signal peptide included in SEQ ID NO:3. Therefore, when SEQ ID NOs:1 and 3 are subjected to global sequence alignment with one another, as mentioned above, amino acids 1-292 of SEQ ID NO:1 align exactly with amino acids 31-322 of SEQ ID NO:3. Reference to particular positions in this specification is by comparison to the positions in full-length NetB SEQ ID NO:3, since the skilled person typically numbers the positions of the full-length protein, rather than the mature truncated protein.

The level of toxicity may be determined as described herein, for example by use of a LMH cell-based LDH assay. The polypeptides of the invention provide protection, when

administered to a subject such as a chicken, from infection by *Clostridium perfringens*. Such protection may be partial, whereby the probability of an individual subject within a population of becoming infected by *C. perfringens* is reduced, or complete, whereby the subject will not become infected by *C. perfringens* (i.e., the probability of becoming infected is 0%).

The term "NetB epitope polypeptide" as used herein means a polypeptide which comprises one or more (or all) epitopes of mature wild-type NetB, as represented by SEQ ID NO:1. The term "epitope" refers to the amino acids (typically a group of around 5 or more amino acids) within a polypeptide sequence which are essential in the generation of an immune response. These amino acids can be consecutive in the sequence but, more typically, are non-consecutive, grouping together when the tertiary structure of the native protein is formed. Provided that these amino acids are within a polypeptide environment which enables them to form the correct epitopic tertiary structure, they can be used to provide a protective vaccine composition. For example, a NetB epitope polypeptide may be one which is capable of binding to an antibody which binds to the mature wild-type NetB having sequence SEQ ID NO:1.

Preferably, the NetB epitope polypeptide comprises a single mutation in one position between 130-297 as compared with the equivalent position in SEQ ID NO:3. Preferably, the mutation comprises one amino acid substitution to an alanine.

The NetB epitope polypeptide may comprise at least 10 contiguous amino acids from SEQ ID NO:29, preferably including at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 46, 47 or 48 contiguous amino acids from SEQ ID NO:30. SEQ ID NO:29 aligns exactly with amino acids 128-195 of SEQ ID NO:3 and SEQ ID NO:30 aligns exactly with amino acids 138-185 of SEQ ID NO:3, when global sequence alignment analysis is used, as mentioned elsewhere herein.

The amino acid sequence shown as SEQ ID NO:30 is a beta-hairpin structure within the NetB protein, which is believed to unfold into the membrane during insertion of NetB into a membrane to form a beta-barrel pore complex. In this embodiment, the mutation in at least one position as compared with the equivalent position in SEQ ID NO:3 is in at least one of positions 130-190 (equivalent to positions 3 to 63 of SEQ ID NO:29), preferably in at least one of positions 138-185 as compared with SEQ ID NO:3 (positions 11-58 of SEQ ID NO:29 and positions 1-48 of SEQ ID NO:30). The polypeptide may comprise a sequence equivalent to SEQ ID NO:30 which differs from SEQ ID NO:30 only in having at least one amino acid substitution, deletion or addition at one or more positions within the sequence, for example at 1-20 positions or 1-10 positions, preferably at 1, 2, 3, 4 or 5 positions. Therefore, the NetB epitope polypeptide of the invention may comprise an amino acid sequence which is at least about 60%, 70%, 80%, 85%, 89%, 91%, 93%, 95% or at least about 97% identical to SEQ ID NO:30, determined using a global sequence alignment comparison as described elsewhere herein.

In one embodiment, the polypeptide may comprise a mutation at either or both of positions P138 and/or position Y182, numbered as found in SEQ ID NO:3. The polypeptide comprising a mutation at position P138 may be non-toxic. In any embodiment, the amino acid P (Proline) at position 138 and/or Y (Tyrosine) at position 182 may be replaced with amino acid A.

Therefore, the polypeptide according to any preceding claim may comprise the amino acid sequence NTISX-EQPDF (SEQ ID NO:25) and/or SYNQNTISXEQP

DFRTIQR (SEQ ID NO:26), where "X" is any amino acid other than Y. Alternatively or additionally, the polypeptide according to any preceding claim may comprise the amino acid sequence ANSIXKNTID (SEQ ID NO:27) and/or NNI-KIANSIX KNTIDKKDVS (SEQ ID NO:28), where "X" is any amino acid other than P. In any of these embodiments, "X" may be amino acid A (Alanine). The polypeptide comprising SEQ ID NO:27 and/or 28 may be non-toxic.

Alternatively, the NetB epitope polypeptide may comprise at least 10 contiguous amino acids from SEQ ID NO:1 and comprising a mutation in at least one position as compared with the equivalent position in SEQ ID NO:3, the polypeptide being capable of binding an antibody which binds to SEQ ID NO:1 and having reduced toxicity compared with the toxicity of SEQ ID NO:1, the polypeptide comprising a β -sandwich domain, a rim domain and a stem domain, wherein the at least one mutation is located in the rim domain. Preferably, the mutation is a conserved residue in the rim domain.

In one embodiment, the NetB epitope polypeptide may comprise at least 10 contiguous amino acids from SEQ ID NO:34, preferably including at least 5, 10, 15, 20, 25, 30, 35, 40, 45, 46, 47 or 48 contiguous amino acids from SEQ ID NO:35. SEQ ID NO:34 aligns exactly with amino acids 212-297 of SEQ ID NO:3 and SEQ ID NO:35 aligns exactly with amino acids 217-292 of SEQ ID NO:3, when global sequence alignment analysis is used, as mentioned elsewhere herein.

The polypeptide may comprise a sequence equivalent to SEQ ID NO:35 which differs from SEQ ID NO:35 only in having at least one amino acid substitution, deletion or addition at one or more positions within the sequence, for example at 1-20 positions or 1-10 positions, preferably at 1, 2, 3, 4 or 5 positions. Therefore, the NetB epitope polypeptide of the invention may comprise an amino acid sequence which is at least about 60%, 70%, 80%, 85%, 89%, 91%, 93%, 95% or at least about 97% identical to SEQ ID NO:35, determined using a global sequence alignment comparison as described elsewhere herein.

In another embodiment, the polypeptide may comprise a mutation at one or more of positions Y221, R230, W287 and/or position W292, numbered as found in SEQ ID NO:3, preferably being a mutation at position W292. In any embodiment, the amino acid at position 221, 230, 287 or 292 may be replaced with amino acid A.

Therefore, the polypeptide may comprise the amino acid sequence YHAIXGNQLF (SEQ ID NO:40) and/or YNIDSYHAIXGNQLFMKSRL (SEQ ID NO:41), where "X" is any amino acid other than Y. Alternatively or additionally, the polypeptide may comprise the amino acid sequence FMKSXLYNNG (SEQ ID NO:42) and/or YGNQLFMKSXLYNNGDKNFT (SEQ ID NO:43), where "X" is any amino acid other than R. In another embodiment, the polypeptide may comprise the amino acid sequence YILNXETTQW (SEQ ID NO: 44) and/or RFDNDYILNXETTQWRGTNK (SEQ ID NO: 45), where X is any amino acid other than W. In a preferred embodiment, the polypeptide comprises the amino acid sequence ETTQXRGTTNK (SEQ ID NO: 46) and/or YILNWETTQXRGTTNKLSSTS (SEQ ID NO: 47), where X is any amino acid other than W. In any of these embodiments, "X" may be amino acid A (Alanine). The polypeptide comprising any of the aforementioned sequences may be non-toxic.

The polypeptide according to the invention may form part of a fusion protein. The polypeptide may have at least about 60% sequence identity to SEQ ID NO: 1, for example, about 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%,

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95%, 96%, 97%, 98% or about 99% sequence identity to SEQ ID NO:1. Sequence identity may be determined using the Needleman-Wunsch Global Sequence Alignment Tool available from the National Center for Biotechnology Information (NCBI), Bethesda, Maryland, USA, for example via blast.ncbi.nlm.nih.gov, using default parameter settings. When comparing the level of sequence identity to SEQ ID NO:1, this typically should be done relative to the whole length of SEQ ID NO:1, to avoid short regions of high identity overlap resulting in a high overall assessment of identity (i.e., a global alignment method is used). For example, a short polypeptide fragment having, for example, five amino acids might have a 100% identical sequence to a five amino acid region within the whole of SEQ ID NO:1, but this does not provide a 100% amino acid identity unless the fragment forms part of a longer sequence which also has identical amino acids at other positions equivalent to positions in SEQ ID NO:1.

An epitope polypeptide according to the invention may be, as mentioned above, any which comprises at least one epitope of NetB and is capable of binding an antibody which will bind to a polypeptide having sequence SEQ ID NO:1. Therefore, the polypeptide may be as little as about 20 amino acids in length provided that it still binds to such an antibody, for example, it may be at least about 30, 40, 50, 70, 90, 120, 150 or about 170 amino acids in length. In some embodiments, the polypeptide may be at least about 190 amino acids in length, for example, it may be between 190 and 360 amino acids in length, such as between 200-350, 220-340 or 250-310 in length. In some embodiments, the polypeptide may be at least about 200 amino acids in length, for example, at least about 220, 230, 240, 250, 260, 270, 280 or about 290 amino acids in length. In certain specific embodiments, the polypeptide may be 292 amino acids in length.

In one embodiment, the polypeptide according to the invention has amino acid sequence SEQ ID NO:5. In another embodiment, it has amino acid sequence SEQ ID NO:6. In further embodiments, the polypeptide according to the invention is selected from one of the amino acid sequences SEQ ID NOs: 36, 37, 38 or 39.

The present invention also encompasses polypeptides comprising variants of the epitope polypeptides and methods utilising these variant polypeptides. As used herein, a "variant" means a polypeptide in which the amino acid sequence differs from the base sequence from which it is derived in that one or more amino acids within the sequence are substituted for other amino acids. The variant is a functional variant, in that the functional characteristics of the polypeptide from which the variant is derived are maintained. For example, the variant polypeptide may have a similar ability to bind an antibody capable of binding to a non-variant polypeptide (such as, by way of non-limiting example, SEQ ID NOs: 5, 6, 36, 37, 38 or 39). In particular, any amino acid substitutions, additions or deletions must not alter or significantly alter the tertiary structure of one or more epitopes contained within the polypeptide from which the variant is derived, so that the variant polypeptide retains the ability to bind to an antibody which binds to SEQ ID NO:1. The skilled person is readily able to determine appropriate functional variants and to determine the tertiary structure of an epitope and any alterations thereof, without the application of inventive skill.

Amino acid substitutions may be regarded as "conservative" where an amino acid is replaced with a different amino

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acid with broadly similar properties. Non-conservative substitutions are where amino acids are replaced with amino acids of a different type.

By "conservative substitution" is meant the substitution of an amino acid by another amino acid of the same class, in which the classes are defined as follows:

Class	Amino acid examples
Nonpolar:	Ala, Val, Leu, Ile, Pro, Met, Phe, Trp
Uncharged polar:	Gly, Ser, Thr, Cys, Tyr, Asn, Gln
Acidic:	Asp, Glu
Basic:	Lys, Arg, His.

As is well known to those skilled in the art, altering the primary structure of a polypeptide by a conservative substitution may not significantly alter the activity of that polypeptide because the side-chain of the amino acid which is inserted into the sequence may be able to form similar bonds and contacts as the side chain of the amino acid which has been substituted out. This is so even when the substitution is in a region which is critical in determining the polypeptide's conformation.

As mentioned above, non-conservative substitutions are possible provided that these do not disrupt the tertiary structure of an epitope within the polypeptide, for example, which do not interrupt the immunogenicity (for example, the antigenicity) of the polypeptide.

Broadly speaking, fewer non-conservative substitutions will be possible without altering the biological activity of the polypeptide. As mentioned above, variants may suitably be at least about 60% identical to the base sequence.

Sequence identity between amino acid sequences can be determined by comparing an alignment of the sequences. When an equivalent position in the compared sequences is occupied by the same amino acid, then the molecules are identical at that position.

Scoring an alignment as a percentage of identity is a function of the number of identical amino acids at positions shared by the compared sequences. When comparing sequences, optimal alignments may require gaps to be introduced into one or more of the sequences, to take into consideration possible insertions and deletions in the sequences. Sequence comparison methods may employ gap penalties so that, for the same number of identical molecules in sequences being compared, a sequence alignment with as few gaps as possible, reflecting higher relatedness between the two compared sequences, will achieve a higher score than one with many gaps. Calculation of maximum percent identity involves the production of an optimal alignment, taking into consideration gap penalties. As mentioned above, the percentage sequence identity may be determined using the Needleman-Wunsch Global Sequence Alignment tool, publicly available via blast.ncbi.nlm.nih.gov, using default parameter settings. The Needleman-Wunsch algorithm was published in J. Mol. Biol. (1970) vol. 48:443-53.

According to a second aspect of the invention, there is provided a polynucleotide having a nucleic acid sequence which encodes for a polypeptide according to the first aspect of the invention, or the complement of such a polynucleotide. Such a polynucleotide may comprise, for example, SEQ ID NOs:31 and/or 32, encoding SEQ ID NOs:5 and 6, respectively or SEQ ID NOs: 48, 49, 50 and/or 51 encoding SEQ ID NOs: 36, 37, 38 and 39 respectively. The invention also encompasses variant nucleic acids encoding the polypeptides of the invention. The term "variant" in relation to

a nucleic acid sequence means any substitution of, variation of, modification of, deletion of, or addition of one or more nucleic acid(s) from or to a polynucleotide sequence, providing the resultant polypeptide sequence encoded by the polynucleotide exhibits at least the same properties as the polypeptide encoded by the basic sequence. The term therefore includes allelic variants and also includes a polynucleotide (a "probe sequence") which substantially hybridises to the polynucleotide sequence of the present invention. Such hybridisation may occur at or between low and high stringency conditions. In general terms, low stringency conditions can be defined as hybridisation in which the washing step takes place in a 0.330-0.825 M NaCl buffer solution at a temperature of about 40-48° C. below the calculated or actual melting temperature (T_m) of the probe sequence (for example, about ambient laboratory temperature to about 55° C.), while high stringency conditions involve a wash in a 0.0165-0.0330 M NaCl buffer solution at a temperature of about 5-10° C. below the calculated or actual T_m of the probe sequence (for example, about 65° C.). The buffer solution may, for example, be SSC buffer (0.15M NaCl and 0.015M tri-sodium citrate), with the low stringency wash taking place in 3×SSC buffer and the high stringency wash taking place in 0.1×SSC buffer. Steps involved in hybridisation of nucleic acid sequences have been described for example in Sambrook et al. (2001; "Molecular Cloning: a laboratory manual", 3rd Edition, Cold Spring Harbor Laboratory Press, New York).

A related aspect of the invention provides a vector comprising a polynucleotide according to the second aspect of the invention and therefore includes recombinant constructs comprising one or more of the nucleic acid molecules described above. The constructs comprise a vector, such as a plasmid or viral vector, into which a nucleic acid molecule of the invention has been inserted, in a forward or reverse orientation. In a preferred aspect of this embodiment, the construct further comprises regulatory sequences, including, for example, a promoter operably linked to the sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art and are commercially available. Appropriate cloning and expression vectors for use with prokaryotic and eukaryotic hosts are also described in Sambrook, et al.

According to a third aspect of the invention, there is provided a cell comprising a polypeptide and/or a polynucleotide and/or a vector according to preceding aspects. For example, a suitable cell may be a *Salmonella* cell, such as a *Salmonella enterica* cell, in some embodiments from the serovar *typhimurium*. The *Salmonella* may be an attenuated strain. Strains χ 8914 and χ 9241 may optionally be employed. Such cells are particularly useful to act as vectors when the polypeptide, polynucleotide and vector of the invention is to be used to provide a vaccine for chickens, to reduce the probability that they will be susceptible to infection by *Clostridium perfringens*. For example, such a system is described in Kulkarni et al. (2008, Vaccine vol. 26: 4194-4203).

According to a fourth aspect of the invention, there is provided a subunit vaccine comprising a polypeptide according to the first aspect of the invention. For example, this may be in the form of a fusion protein and/or in the form of a recombinant viral vaccine.

A fifth aspect of the invention provides a vaccine composition comprising a polypeptide and/or a polynucleotide and/or a vector and/or a cell and/or a subunit vaccine according to preceding aspects of the invention. The composition may further comprise excipients and/or diluents

appropriate for the means by which the composition is to be administered to a subject in need of vaccination against infection by *C. perfringens*. Selection of appropriate components is within the routine capability of the skilled person without the application of inventive activity.

For example, the vaccine composition of the invention may conveniently be formulated using a pharmaceutically acceptable excipient or diluent, such as, for example, an aqueous solvent, non-aqueous solvent, non-toxic excipient, such as a salt, preservative, buffer and the like. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oil and injectable organic esters such as ethyl oleate. Aqueous solvents include water, alcoholic/aqueous solutions, saline solutions, parenteral vehicles such as sodium chloride, Ringer's dextrose, etc. Preservatives include antimicrobial, anti-oxidants, chelating agents and inert gases. The pH and exact concentration of the various components the vaccine composition are adjusted according to routine skills.

Optionally, the vaccine formulation may include a carrier. Commonly used carrier molecules are bovine serum albumin (BSA), keyhole limpet hemocyanin (KLH), ovalbumin, mouse serum albumin, rabbit serum albumin and the like. Synthetic carriers may be used and are readily available. Means for conjugating peptides to carrier proteins are well known in the art and include glutaraldehyde, m-maleimido-benzoyl-N-hydroxysuccinimide ester, carbodiimide and bis-biotinized benzidine.

In certain situations, it may also be desirable to formulate the vaccine composition to comprise an adjuvant to enhance the immune response. Such adjuvants include all acceptable immunostimulatory compounds such as, for example, a cytokine, toxin, or synthetic composition. Commonly used adjuvants include aluminium hydroxide, aluminium phosphate, calcium phosphate, Freund's adjuvants and Quil-A saponin. In addition to adjuvants, it may be desirable to co-administer biologic response modifiers (BRM) with the peptide or variant or derivative to down regulate suppressor T cell activity.

Possible vehicles for administration of the vaccine formulation include liposomes. Liposomes are microscopic vesicles that consist of one or more lipid bilayers surrounding aqueous compartments. Liposomes are similar in composition to cellular membranes and, as a result, liposomes generally can be administered safely and are biodegradable. Techniques for preparation of liposomes and the formulation (e.g., encapsulation) of various molecules, including peptides and oligonucleotides, with liposomes are well known.

Depending on the method of preparation, liposomes may be unilamellar or multilamellar and can vary in size with diameters ranging from 0.02 μ m to greater than 10 μ m. Liposomes can also adsorb to virtually any type of cell and then release the encapsulated agent. Alternatively, the liposome fuses with the target cell, whereby the contents of the liposome empty into the target cell. Alternatively, an absorbed liposome may be endocytosed by cells that are phagocytic. Endocytosis is followed by intralysosomal degradation of liposomal lipids and release of the encapsulated agents. In the present context, the polypeptide according to the invention can be localized on the surface of the liposome, to facilitate antigen presentation without disruption of the liposome or endocytosis. Irrespective of the mechanism or delivery, however, the result is the intracellular disposition of the associated polypeptide.

Liposomal vectors may be anionic or cationic. Anionic liposomal vectors include pH sensitive liposomes which disrupt or fuse with the endosomal membrane following

endocytosis and endosome acidification. Cationic liposomes are preferred for mediating mammalian cell transfection in vitro, or general delivery of nucleic acids, but are used for delivery of other therapeutics, such as peptides.

Other suitable liposomes that are used in the methods of the invention include multilamellar vesicles (MLV), oligolamellar vesicles (OLV), unilamellar vesicles (UV), small unilamellar vesicles (SUV), medium-sized unilamellar vesicles (MIN), large unilamellar vesicles (LUV), giant unilamellar vesicles (GUV), multivesicular vesicles (MVV), single or oligolamellar vesicles made by reverse-phase evaporation method (REV), multilamellar vesicles made by the reverse-phase evaporation method (MLV-REV), stable plurilamellar vesicles (SPLV), frozen and thawed MLV (FATMLV), vesicles prepared by extrusion methods (VET), vesicles prepared by French press (FPV), vesicles prepared by fusion (FUV), dehydration-rehydration vesicles (DRV), and bubblesomes (BSV). Techniques for preparing these liposomes are well known in the art.

Other forms of delivery particle, for example, microspheres and the like, also are contemplated for delivery of the peptide epitopes or polypeptides.

Alternatively, nucleic acid-based vaccines may be produced that comprise nucleic acid, such as, for example, DNA or RNA, encoding the immunologically active peptide epitope or polypeptide and cloned into a suitable vector (e.g., vaccinia, canarypox, adenovirus, or other eukaryotic virus vector).

Alternatively, the polypeptide may be administered in the form of a cellular vaccine via the administration of autologous or allogeneic APCs or dendritic cells that have been treated in vitro so as to present the peptide on their surface. *Salmonella* cells may also be used, especially for administration to chickens. This involves the use of live attenuated *Salmonella* vaccines to deliver the antigen. This approach offers a number of advantages. First, live *Salmonella* vaccines can be given orally (the natural route of infection), enabling a non-invasive route of vaccine administration. Second, both mucosal and systemic immune responses can be elicited, which may be important for protection against infection. In addition, live attenuated *Salmonella* vaccines are able to simulate both humoral and cellular immune responses that may be important for protection against disease. Finally, since *Salmonella* is genetically tractable, recombinant *Salmonella* vaccines are relatively easy to develop and are also relatively cost effective to produce.

One of the most widely studied classes of attenuated *Salmonella* used as carriers of foreign antigens are auxotrophs. For example, genetically defined mutants of the *aroA* gene, encoding 5-enolpyruvylshikimate-3-phosphate synthase, have been constructed in both *S. enterica* var. Typhimurium and var. Typhi. These mutants are attenuated and immunogenic in mice. Examples of other auxotrophic mutants include *Salmonella* with deletions in the genes involved in the purine biosynthetic pathway. Another well-studied group of attenuated *Salmonella* are mutants that have defined deletions in genes involved in the regulation of *Salmonella* virulence. For example, mutations in genes encoding adenylate cyclase (*cya*) and camp receptor protein (*crp*) affect the expression of genes involved.

In one embodiment, the vaccine composition may be included in an animal feed (i.e., a foodstuff suitable for consumption by an animal, particularly a chicken) comprising a polypeptide and/or a polynucleotide and/or a vector and/or a cell and/or a subunit vaccine and/or vaccine composition according to preceding aspects of the invention. This may, in non-limiting examples, be in the form of

pellets, crumbs or a mash which may further comprise, again for example only, grain, grass and/or protein components. The composition may also be included in drinking liquids and/or administered via a spray into the atmosphere surrounding the animal which is, consequently, inhaled by the animal.

In a sixth aspect of the invention, a polypeptide and/or a polynucleotide and/or a vector and/or a cell and/or a subunit vaccine and/or a vaccine composition according to any preceding aspect is for use in a method of vaccinating a subject against infection by *Clostridium perfringens*.

Likewise, a seventh aspect of the invention provides a method of vaccinating a subject against infection by *Clostridium perfringens* comprising administering to the subject a protective amount of a polypeptide and/or a polynucleotide and/or a vector and/or a cell and/or a subunit vaccine and/or a vaccine composition according to any preceding aspect. A "protective amount" is an amount sufficient to induce an immune response in the subject, such that the probability of the subject becoming infected by *C. perfringens* if exposed to the bacterium is reduced or removed. For example, antibodies capable of binding to SEQ ID NO:1 may be detectable after the administration, where such antibodies were not detectable prior to the administration, or only detectable at lower concentrations than after administration.

In the sixth and seventh aspects, the subject may be of the genus *Gallus*, for example, of the species *Gallus gallus* (i.e., the domestic chicken). When the subject is a chicken, the preferred means for delivery of the polypeptide and/or a polynucleotide and/or a vector and/or a cell and/or a subunit vaccine of the other aspects of the invention may be a *Salmonella*-based system as described herein. The subject may also be a mammalian subject, for example, a human.

According to an eighth aspect of the invention, there is provided a kit comprising a polypeptide and/or a polynucleotide and/or a vector and/or a cell and/or a subunit vaccine and/or a vaccine composition according to any of the preceding aspects. For example, the kit may be a kit for use by a veterinarian or farmer to vaccinate a flock of chickens and may comprise a *Salmonella* vector comprising a polypeptide according to the invention, for example for administration to chickens by inclusion in their feed.

BRIEF DESCRIPTION OF FIGURES

Embodiments of the invention will be described, by way of example only, with reference to the following FIGS. 1-12 in which:

FIG. 1 shows an SDS PAGE gel of a purified recombinant NetB (rNetB) in lane 2, with lane 1 being marker (molecular mass is indicated in kDa at the site);

FIG. 2 shows a Western Blot of rNetB with α -Xpress antibodies in lane 2, with lane 1 being marker (molecular mass is indicated in kDa at the site);

FIG. 3 shows phase-contrast microscopy images showing morphological damage of LMH cells induced by incubation with rNetB, with panels A and B being control cells and panels C and D showing cells exposed to rNetB (7.7×10^{-10} mol, 1 h), in which cell swelling and cell blebbing induced by rNetB can be observed;

FIG. 4 shows the cytotoxic effect of different concentrations of rNetB on LMH cells;

FIG. 5 shows an SDS PAGE gel of formaldehyde derived NetB toxoid in lane 2, with lane 1 being marker (molecular mass is indicated in kDa at the site);

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FIGS. 6A, 6B and 6C illustrate the crystal structure of NetB, with FIG. 6A being a ribbon representation of an isolated NetB subunit, FIG. 6B being a close up view of the rim domain 2 and FIG. 6C being a ribbon representation of the NetB assembly viewed from the side;

FIGS. 7A to 7D illustrate a functional analysis of various NetB rim mutants;

FIG. 8 shows a necrotic enteritis model for studying immunisation of broiler chickens with antigen over time;

FIG. 9 shows an SDS PAGE gel of a purified NetB mutant W292A in lane 2, with lane 1 being marker, the arrows indicating monomeric and heptameric forms of NetB (molecular mass is indicated in kDa at the site);

FIG. 10 shows an SDS PAGE gel of wild type NetB in lane 2, with lane 1 being marker, the arrows indicating monomeric and heptameric forms of NetB (molecular mass is indicated in kDa at the site);

FIG. 11 shows the cytotoxic effect of wild type NetB (panel B), formaldehyde NetB toxoid (panel C) and NetB W262A (panel D) on LMH cells (panel A shows untreated cells); and

FIG. 12 shows the lesion scores of individual animals (top) and percentage NE positive chickens (bottom) after vaccination of animals with a formaldehyde NetB toxoid or NetB W262A.

EXAMPLES

Bacterial Strains and Plasmids

Bacterial strains and plasmids used in this study are listed in Table 1.

TABLE 1

Bacterial strains and plasmids used in this study		
Bacterial strains and plasmids	Relevant genotype or phenotype	Source
<i>E. coli</i> TOP10	F- mcrA Δ(mrr-hsdRMS-mcrBC) Φ80lacZAM15 ΔlacX74 recA1 araD139 Δ(araleu) 7697 galU galK rpsL (Str ^R) endA1 nupG	Invitrogen
<i>C. perfringens</i> strain 56	wild-type, virulent NE isolate	(Gholamiandekhordi et al. (2006) <i>Vet Microbiol</i> vol 113: 143-152)
pCR4-TOPO	kan, bla	Invitrogen
pCR4-TOPO-netB	netB	This study
pBAD	araBAD promoter, bla	Invitrogen
pBAD-netB	netB	This study

The pBAD vector (Invitrogen) was used as an expression vector and *E. coli* Top10 (Invitrogen) was used as an expression host. The pCR4-TOPO cloning vector (Invitrogen) was used as an intermediate vector to facilitate cloning the genes of interest. *C. perfringens* strain 56 (Gholamiandekhordi et al. (2006) *Vet Microbiol* vol 113: 143-152) was used for amplification of the netB gene. *C. perfringens* was grown anaerobically at 37° C. in brain heart infusion (BHI) broth. *E. coli* strains were grown either in terrific broth (TB) or Luria-Bertani (LB) agar supplemented with ampicillin (100 µg/ml) and grown at 37° C.

Molecular Cloning of netB

Bacterial genomic DNA from *C. perfringens* strain 56 was isolated with the Wizard Genomic Purification Kit (Promega) and used as a template to amplify the encoding

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region of the netB gene (GenBank accession number: A8ULG6; SEQ ID NO:4) using the following primers:

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netB_fwd:
5' -CCCGGGCTCGAGAGTGAATTAATGACATAAAC-3' (SEQ ID NO: 7)

netB_rev:
5' -CCCGGGAAGCTTTTACAGATAATATTCTATTTATG-3' (SEQ ID NO: 8)

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Thereby, the netB was amplified from SEQ ID NO:33, lacking the N-terminal signal sequence (30 amino acids). XhoI and HindIII were used as restriction endonucleases and included in the primers for subcloning into the pBAD bacterial expression vector (underlined in the sequences above). The amplified fragments were first cloned into a pCR4-TOPO cloning vector and verified by sequencing. These fragments were then cloned into the pBAD vector to generate recombinant pBAD-netB and used to transform *E. coli* TOP10 competent cells. Thereby, the recombinant netB is expressed as a 331 amino acids protein (SEQ ID NO:2, having a calculated molecular mass of 38kDa), containing the mature NetB protein (SEQ ID NO:1) without the native signal peptide and also containing a His-tag and a Xpress motif for purification and specific detection of the protein. Expression and Purification of Recombinant NetB in *E. coli*

The recombinant *E. coli* carrying the pBAD-netB vector was grown to an optical density (OD) of 0.5 and expression was induced for 6 h by adding arabinose at a final concentration of 0.02%. Bacterial cells were harvested by centrifugation and cell lysis was achieved by using BugBuster reagent (Novagen). Recombinant NetB protein was purified by His-bind chromatography columns (GE Healthcare) according to the manufacturer's instructions. Subsequently, the protein was dialysed into Tris-buffered saline (TBS) by using PD-10 desalting columns (GE Healthcare) and protein concentrations were measured with a UV-Vis spectrophotometer (Pierce).

SDS-PAGE and Western Blotting

The protein yield and purity was analysed by SDS-PAGE on precast 4-12% acrylamide-bisacrylamide gels (Novagen). Therefore, gels were run in MES running buffer (Novagen) for about 45 min at 200V and stained with SimplyBlue Safestain (Invitrogen). The perfect protein marker (Novagen) was used as a protein standard. Protein specificity was determined by Western blotting. Therefore, proteins were transferred after electrophoretic separation onto a nitrocellulose membrane with an iBlot dry blotting system (Invitrogen) and blots were developed using primary anti-Xpress specific antibodies (Invitrogen) followed by a secondary anti-mouse antibody conjugated to horseradish peroxidase (HRP). Visualisation was performed with the Amersham ECL Western blotting detection reagents (GE Healthcare) and photographic films were exposed to varying time periods. The precision plus protein WesternC (Biorad) was used as a protein standard.

The results are shown in FIG. 1 (SDS-PAGE) and FIG. 2 (Western blot). The images show monomeric (≈38 kDa) and heptameric (≈266 kDa) forms of the protein.

Cytotoxicity Assay for Effect of rNetB on LMH Cells

Cytotoxicity was measured using the CYTOTOX96® kit (Promega), in which cytotoxicity is measured as the amount of LDH (lactate dehydrogenase) released from the cell cytosol into the medium caused by the presence of the tested pore-forming cytotoxic compound. The rNetB was evaluated for its cytotoxicity by incubation with chicken hepatocellular carcinoma (LMH; ATCC: CRL-2117) cells, a cell line known to be susceptible for the toxin.

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Therefore, LMH cells were grown in Waymouth's MB 752/1 medium (Invitrogen) supplemented with 10% fetal calf serum at 37° C. in a 5% CO₂ incubator to 70-80% confluency in 96-well plates. FIG. 3 shows the morphological effects of rNetB on LMH cells. Control cells (FIGS. 3A and 3B) demonstrate the epithelial and dendritic-like growth of LMH cells in cell culture. Treatment of cells with purified rNetB (7.7⁻¹⁰ mol, 1 h) caused rapid cell blebbing and cell swelling (FIGS. 3C and 3D). Longer incubation periods lead to total cell lysis (data not shown).

Consequently, cells were incubated with serial dilutions of NetB in Waymouth's medium (100 µl final volume in each well) for 2 h at 37° C. Control cells were incubated with Waymouth's medium to determine either the base line (0%) or total cell lysis (100%), achieved by freezing and thawing of the cells. After 2 h of incubation the supernatant was assayed and percentage cytotoxicity was determined relative to the control groups (FIG. 4). Each dilution was assayed in six replicates and in three independent experiments (data are means±standard deviations). The median cytotoxic dose (CT₅₀) was determined as 29 µg/ml (7.7⁻¹⁰ mol). Generating Formaldehyde Derived Netb Toxoid as Candidate Vaccine

The recombinant NetB was suspended in Tris-buffered saline (TBS) at 400 µg/ml and formaldehyde added to a final concentration of 130 mM. After incubation for 5 days at 37° C., the reaction was stopped by the addition of L-Lysine to 30 mM final concentration and residual formaldehyde was removed by dialysis against TBS buffer. Formaldehyde treatment of the rNetB led to a highly cross-linked protein (FIG. 5). The formaldehyde-derived NetB toxoid was incubated with LMH cells and no cytotoxic effect could be observed (data not shown).

Construction of NetB Mutants

In order to map key residues critical for NetB functionality (cell binding, oligomerisation, pore-formation) a monomeric/heptameric protein model of NetB has been made based on sequence similarities with related pore-forming toxins (data not shown). As a result, the following residues were selected to be replaced by an alanine by site-directed mutagenesis: D81, P138, Y153, G157, Y182, Q184, P185 and R230. The mutants were constructed with the QuikChange II site-directed mutagenesis Kit (Stratagene) by using the primers listed in Table 2 below.

The recombinant pBAD-netB expression vector was then used as a template to amplify the respective mutant netB gene. The rNetB mutants were expressed and purified as described earlier for the rNetB but only two mutants (P185A, R230A) behaved as the wt rNetB in terms of protein stability. The other rNetB mutants (D81A, P138A, Y153A, G157A, Y182A, Q184A) were less soluble and as a consequence were mainly accumulated in inclusion bodies during protein expression. Although in low protein amounts and not very pure, the less stable rNetB mutants could be purified and preliminary data from incubating the rNetB mutants with LMH cells suggest a less-toxic (Y182A) and a non-toxic mutant (P138A).

TABLE 2

Primers used in this study for netB mutagenesis		
Primer	Sequence (5'-3')	SEQ ID NO
netB_D81A_fwd	GGAACATTATTGAAGCTCCTCAT TCTGATAAGAACTGC	9

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TABLE 2-continued

Primers used in this study for netB mutagenesis		
Primer	Sequence (5'-3')	SEQ ID NO
netB_D81A_rev	GCAGTTTCTTATCAGAATGAGGA GCTTCAATAAATGTTCC	10
netB_P138A_fwd	GCAAATCTATTGCTAAAAATACT ATAGATAAAAAGATGTATC	11
netB_P138A_rev	GATACATCTTTTATCTATAGTA TTTTTAGCAATAGAATTGTC	12
netB_Y153A_fwd	GATGTATCTAATTCAATTGGTGGC TCTATAGGCGG	13
netB_Y153A_rev	CCGCCTATAGACGCCCAATTGAA TTAGATACATC	14
netB_G157A_fwd	CAATTGGTTATTCTATAGGCGCTA ATATATCTGTTGAAGG	15
netB_G157A_rev	CCTTCAACAGATATATTAGCGCCT ATAGAATAACCAATTG	16
netB_Y182A_fwd	GTCCAAAATACTATAAGCGCTGAA CAACCTGATTTTAGAAC	17
netB_Y182A_rev	GTTCTAAAATCAGGTTGTTACGCG CTTATAGTATTTTGAC	18
netB_Q184A_fwd	CCAAAATACTATAAGCTATGAAGC ACCTGATTTTAGAACCAATTC	19
netB_Q184A_rev	GAATTGTTCTAAAATCAGGTGCTT CATAGCTTATAGTATTTTGG	20
netB_P185A_fwd	CTATAAGCTATGAACAAGCTGATT TTAGAACAAATTCAAAG	21
netB_P185A_rev	CTTTGAATTGTTCTAAAATCAGCT TGTTTCATAGCTTATAG	22
netB_R230A_fwd	CAATTATTCATGAATCAGCATTG TATAATAATGGTG	23
netB_R230A_rev	CACCATTATTATACAAATGCTGATT TCATGAATAATTG	24

Expression of rNetB Polypeptides in *Salmonella*

The polynucleotide encoding a NetB polypeptide is expressed in an attenuated *Salmonella* strain such as *Salmonella enterica* serovar *Typhimurium* (e.g., strain SL2361 or χ 9241 or χ 9352 or AviPro *Salmonella* Vac T), *Salmonella enterica* serovar *Enteritidis* (e.g. AviPro *Salmonella* Vac E), or *Salmonella enterica* serovar *Gallinarum* (e.g. strain JOL916 or Gallivac). A number of different approaches are used. The NetB polynucleotide is cloned into a plasmid (for example, plasmid pSC1901, pSEC10 or pBR322), or inserted onto the chromosome of the *Salmonella* strain (for example into a gene in the shikimate pathway). The expression of the NetB polynucleotide is driven by a constitutive or an inducible promoter (for example the phoP or ompC gene promoter). The protein is exported by fusing it to a component of a system such as ClyA.

Vaccination in a Mouse Model

A recombinant *Salmonella* vaccine is tested in BALB/c mice. Groups of 10 mice are immunised intragastrically using a gavage needle with approximately 10⁹ cfu of recombinant *Salmonella* expressing at least one rNetB mutant as described above, for example, Y182A comprising polypep-

tide sequence SEQ ID NO:5 or P138A comprising polypeptide sequence SEQ ID NO:6. The cells are re-suspended in 100 µl LB broth. Mice are immunised on days 0, 14, 28, 42 and 56. Tail vein serum samples are collected on days 13, 27, 41, 55, 69 and 83 and the concentration of any antibody against NetB present determined using an ELISA.

Vaccination in a Chicken Model

20 µg of toxoid is administered with QuilA adjuvant (1:1) to each bird. Three doses are given at one week intervals, intramuscularly in the breast muscle. Animals are bled at 1 week after the last immunisation and challenge and the concentration of any antibody against NetB present determined using an ELISA. For mass dosing the vaccine is administered via drinking water, foodstuffs or as a spray. Crystallisation and Data Processing.

The NetB gene (GenBank accession number: A8ULG6) was cloned, overexpressed in *E.coli* and purified in monomeric form using Ni-NTA chromatography. NetB oligomer was obtained by incubating monomeric NetB with cholesterol rich liposomes and subsequently solubilised using detergent.

NetB oligomer was crystallised initially using the sitting drop method in 12% (w/v) PEG 4000, 100 mM sodium cacodylate pH 6.5. However, these crystals were too small for diffraction studies. Optimisation using a combination of limited trypsin proteolysis and the addition of the additive detergent polyethylene glycol dodecyl ether (THESIT[®] part of the Hampton detergent screen) led to crystals which diffracted to ~4 Å. Data were collected on a Rigaku Saturn 944 CCD detector mounted onto a Rigaku Micromax X-ray generator. Data were indexed and integrated with Mosflm (Battye TGG et al (2011) *Acta Crystallographica Section D* vol 67(4): 271-281), and scaled using SCALA (Evans P. (2006) *Acta Crystallogr D Biol Crystallogr.* 62 (Pt 1): 72-82). Molecular replacement was carried out using Phaser MR (McCoy et al. (2007) *J Applied Crystallogr.* 40 (Pt 4): 658-674) and the αHL heptamer structure (PDB ID 7AHL) as a search model. The initial model refined using Phenix (Adams PD et al. (2010) *Acta Crystallogr D Biol Crystallogr.* 66 (Pt 2): 213-221) with NCS, Ramachandran and secondary structure restraints, and increased weighting on stereochemical terms. At the final stages, refinement was carried out using Buster (Smart OS, et al. (2012) *Acta Crystallogr D Biol Crystallogr.* 68 (Pt 4): 368-380) with NCS and reference structure restraints (αHL used as a closely related reference structure). Manual building and real space refinement was performed using COOT (Elmsley Petal. (2010) *Acta Crystallogr D Biol Crystallogr.* 66 (Pt 4): 486-501), and model validation was calculated using Molprobity (Davis et al. (2007) *Nucleic Acids Res* 35: W375-383). The PYMOL[™] Molecular Graphics System was used for visualisation and electrostatic potential surface rendering. A model with all sidechains present was used for electrostatic potential calculation even when side-chain density was not visible in the electron density map.

The crystal structure of NetB is illustrated in FIGS. 6A to 6C of the accompanying drawings. FIG. 6A is a ribbon representation of an isolated NetB subunit. As with αHL, there are three domains; a β-sandwich domain 1, a rim domain 2 and a stem domain 3. FIG. 6B is a close up view of the rim domain 2. Residues in NetB that were further investigated by mutation to evaluate their function are shown in stick representation. FIG. 6C illustrates the NetB assembly viewed from the side.

Each NetB monomer contains 15 β-strands accounting for 53% of the polypeptide, whilst the remainder is made up of one 3₁₀-helix, one short α-helix and random coil.

The β-sandwich domain consists of two β-sheets composed of strands 1, 2, 3, 6, 11 and 5, 9, 10, 12, 14, 15, respectively, and a single α-helix. Strands 5 and 12 extend into the lower part of the molecule, and these strands, along with strands 4 and 13, the 3₁₀-helix and considerable random coil, make up the rim domain. The stem domain contains the long, curved amphipathic hairpin (strands 7 and 8), which is connected to the β-sandwich domain through two short coils, forming a triangle region similar to that in other membrane-inserted βPFTs. Superposition with the *S. aureus* αHL monomer results in an rmsd of 1.55 Å. With the exception of two small β-strands which are absent in NetB (αHL strands 11 and 12), secondary structure elements are largely conserved between the two toxins. Areas of small structural deviation to αHL exist in all three domains. For example, loop positions in the β-sandwich domain between strands 5 and 6 and strands 11 and 12 deviate from the corresponding loops in αHL. Strand 15 in αHL, which spans the rim and β-sandwich domains, is composed of two separate strands (13 and 14) in NetB. Finally, while the NetB stem domain adopts a similar curvature to αHL, the turn at the cytoplasmic end bends inwards towards the pore lumen in NetB. The most interesting differences between the two toxins lie in the area that constitutes the rim loops.

The rim domain is thought to be the region that interacts with the target cell membrane. Therefore, further studies were carried out to determine whether mutation of key residues along the rim loops, affect NetB function. Mutants showing a significant difference relative to wild type NetB are summarised in Table 3 below.

FIGS. 7A to 7D show binding ability of various mutants to LMH cells relative to wild type NetB. LMH cytotoxicity was performed as outlined hereinafter. Mutants Y191A, R200A, W257A and W262A showed significantly reduced binding compared to wild type toxin, see FIG. 7A. Subsequently, the three NetB mutants showing the lowest affinity in the ON CELL WESTERN[™] assay (Y191A, R200A, and W262) were tested for their abilities to cause lysis of LMH cells at 4, 8, or 16 µM concentrations (FIG. 7B). All 3 mutants showed reduced cytotoxicity relative to wild type NetB, with W262A showing the highest decrease.

TABLE 3

Functional characteristics of NetB mutants.			
NetB mutant	Reduced binding on LMH cells	Reduced haemolysis on RBCs	Reduced cytotoxicity on LMH cells
Y78A	/	***	N.D.
Y187A	/	**	N.D.
H188A	/	***	N.D.
Y191A	**	***	*
R200A	**	***	**
Y202A	/	***	N.D.
W257A	*	***	N.D.
W262A	***	***	***

(Asterisks indicate a statistically significant difference relative to wild type NetB (***: p < 0.001; **: p < 0.01; *: p < 0.05; 1-way ANOVA). N.D.: not determined).

In addition, the hemolytic activity of NetB was evaluated by incubating the toxin with human red blood cells (hRBCs). hRBC hemolysis was performed using standard procedure known in the art. FIG. 7C shows the CT₅₀ values of the NetB variants. However, CT₅₀ values of mutants Y191A, R200A, W257A, and W262A could not be determined as they only caused incomplete hemolysis within 1 h, even at 5 µM concentrations. Therefore, these mutants were incubated with hRBCs for an extended period and degree of hemolysis was monitored at 2, 4, and 24 h (FIG. 7D). Again, mutants

Y191A, R200A, W257A and W262 showed the most significant decrease in activity relative to wild type NetB. In addition, mutants Y78A, Y187A, H188A and Y202A also showed a significant increase in CT_{50} values.

In summary, replacement of conserved residues along the rim loops of NetB (Y191, R200, W257, and W262) had the most dramatic effect on NetB cell binding and toxicity. In addition, due to the broader dynamic range of the hemolysis assay, it could be shown that non-conserved residues such as Y78, Y187, H188, and Y202 also play a role in NetB function.

The position of the mutations referred to in Table 3 above is numbered as found in SEQ ID NO:1 and thus the position of the mutation in the full-length protein will be plus 30 amino acids. Accordingly, Y191A is Y221A (SEQ ID NO:36), R200A is R230A (SEQ ID NO: 37), W257A is W287A (SEQ ID NO: 38) and W262A is W292A (SEQ ID NO: 39). The NetB mutant W262A (W292A in SEQ ID NO:3) was shown to cause the most significant reduction in toxicity so it was decided to test NetB W262A (SEQ ID NO: 39) or a formaldehyde NetB toxoid for their potential to protect chickens against NE in an in vivo model.

The expression and purification of NetB in *E.coli* was carried out as hereinbefore described using the bacterial strains and plasmids listed in Table 1. The preparation of a formaldehyde NetB toxoid was also carried out as outlined above.

Expression and purification of NetB W262A(W292A) in *E.coli*.

Bacterial cells were grown in TB at 37° C. and expression of NetB induced by the addition of arabinose. The cells were lysed using BugBuster and NetB purified using a His-tag column at a concentration of 1.7 mg/ml.

The purified proteins were analysed by SDS-PAGE as previously described and the results are shown in FIGS. 9 and 10.

Cytotoxicity Towards LMH Cells.

The NetB was evaluated for its cytotoxicity to a chicken hepatocellular carcinoma epithelial cell line (LMH; ATCC: CRL-2117). Therefore, LMH cells were grown in Waymouth's MB 752/1 medium (Invitrogen) supplemented with 10% fetal calf serum at 37° C. in a 5% CO₂ incubator to 70-80% confluency on 96-well plates. Subsequently, cells were incubated with 0.4 mg/ml of wild type NetB, formaldehyde NetB toxoid, or NetB W262A. Effects on cell morphology were observed with an optical microscope, as shown in FIG. 11. Untreated cells had epithelial and dendritic-like growth (panel A). Treatment of the cells with purified NetB caused rapid cell blebbing and swelling (panel B). In contrast, incubation with a formaldehyde NetB toxoid or NetB W262A did not result in morphological changes indicative of toxicity to LMH cells (panels C and D). Immunisation with Formaldehyde NetB Toxoid and NetB W262A.

A formaldehyde NetB toxoid and NetB W262A were used in an in vivo model. Ross 308 broiler chickens were obtained as one-day-old chicks from a local commercial hatchery. All treatment groups were housed in the same room. The birds were reared in pens at a density of 25 animals per 1 m² on wood shavings. All pens were separated by solid walls to prevent contact between birds from different treatment groups. Before the trial, the rooms were decontaminated with Metatectyl HQ (CLIM'OMEDIC®, Metatecta, Belgium) and a commercial anticoccidial desinfectant (OOCIDE, DuPont Animal Health Solutions, Wilmington, US). The chickens were divided in 4 groups of 25 animals. They received ad libitum drinking water and feed.

A 23 h/1 h light/darkness program was applied. The animal experiments were carried out according to the recommendations and following approval of the Ethical Committee of the Faculty of Veterinary Medicine, Ghent University.

The NE model was based on the subclinical in vivo model described previously in Gholamiandehkordi et al (2007) *Avian Pathol* October; 36(5): 375-82. Groups of 25 one-day-old Ross 308 broiler chickens were fed a wheat/rye-based (43%/7.5%) diet, with soybean meal as protein source. The feed composition is described in Gholamiandehkordi et al (supra). Nobilis Gumboro D 78 vaccine (Schering-Plough Animal Health, Brussels, Belgium) was given in the drinking water on day 16 in all groups. From day 17 onwards soy bean meal was replaced by fishmeal (30%) as protein source. All groups were orally challenged once a day on day 17, 18, 19 and 20 with approximately 4×10^8 cfu *C. perfringens* strain 56 bacteria. On day 18 all groups were orally inoculated with a ten-fold dose of Paracox-5 (Schering-Plough Animal Health, Brussels, Belgium). On day 21, 22, 23, each time one-third of the birds was euthanized and necropsied, as schematically shown in FIG. 8.

Lesions in the small intestine (duodenum to ileum) were scored (as described by Keyburn et al. (2006) *Infect Immun* vol 74: 6496-6500) as follows: 0=no gross lesions; 1=congested intestinal mucosa; 2=small focal necrosis or ulceration (1-5 foci); 3=focal necrosis or ulceration (6-15 foci); 4=focal necrosis or ulceration (16 or more foci); 5=patches of necrosis 2-3 cm long; 6=diffuse necrosis typical of field cases. Lesion scores of 2 or more were classified as NE positive. As controls, animals were unimmunized or treated only with adjuvant.

As demonstrated in FIG. 12, both antigens significantly reduced lesion scores relative to the control groups. No difference could be detected between the non-vaccinated and the adjuvant-only controls. This clearly shows that a formaldehyde NetB toxoid or NetB W262A could also form the basis of an effective vaccine for NE. Although both antigens significantly increased protection, the usage of a recombinant protein brings the added advantage of not having to test every batch of a formaldehyde NetB toxoid for safety.

Discussion

In this study, an expression system for netB in *E. coli* has been established and it has been shown that the recombinant protein is able to form oligomeric complexes. Cytotoxicity assays on LMH cells, a cell line approved to be susceptible for NetB (Keyburn et al., (2008) *PLoS Pathog* vol 4: e26), showed that the recombinant toxin is capable of forming functional pores and causing cell lysis. Based on sequence similarities with related pore-forming toxins, a monomeric/heptameric model of NetB has been designed (data not shown) and used to identify amino acids that, if replaced by another, could have a dramatic impairment on NetB functionality, e.g. cell binding, oligomerisation, or pore-formation. Consequently, eight NetB mutants were designed by site directed mutagenesis (D81A, P138A, Y153A, G157A, Y182A, Q184A, P185A, R230A).

Surprisingly, preliminary incubation of these rNetB mutants on LMH cells indicate that only two of the mutants had reduced toxicity, with one less-toxic Y182A and one non-toxic (P138A) mutant (compared to wild type rNetB).

The tyrosine at position 182 is located within the cap region and might be crucial maintaining the correct orientation of the stem and the cap, as well as for interacting with the membrane-lipids during pore-formation. The proline at position 138 is thought to play a crucial role in the pore-formation process, as it is located at the end of the beta-hairpin structure. During pore-formation, the unfolding of

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the beta-hairpin into the membrane is an essential step in building up the functional beta-barrel pore complex.

This study has used a formaldehyde-derived toxoid or a non-toxic variant of the NetB to immunise chicken and thereby stimulating a specific antibody response to protect chicken from a subsequent toxin challenge.

As described herein, further studies were then carried out to investigate the crystal structure of the heptameric complex of NetB in detergent. The heptameric structure, which is likely to represent the membrane-inserted pore-form, was found to have high structural similarity to the Staphylococcal toxin α -HL, revealing conservation of many of the key residues that are important for function in this family of β -PFTs but displaying differences that may have evolved separately in the Clostridial counterparts. Residues critical for NetB binding and toxicity were also identified.

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As described above, replacement of conserved residues along the rim loops of NetB (Y191, R200, W257, and W262) had the most dramatic effect on NetB cell binding and toxicity. In addition, due to the broader dynamic range of the hemolysis assay, it could be shown that non-conserved residues such as Y78, Y187, H188, and Y202 also play a role in NetB function.

Thus it can be seen that mutation of residues contained within the rim domain, particularly those highly conserved in β -PFTs of *S.aureus* and *C. perfringens* or within a pore-forming domain, such as the β -hairpin structure can significantly affect host-cell binding and consequently, cytotoxicity of NetB. Such recombinant proteins have real potential for use as an effective vaccine against NE.

SEQUENCE LISTING

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Thr Thr Gln Trp Arg Gly Thr Asn Lys Leu Ser Ser Thr Ser Glu Tyr	
290 295 300	
Asn Glu Phe Met Phe Lys Ile Asn Trp Gln Asp His Lys Ile Glu Tyr	
305 310 315 320	
Tyr Leu	

<210> SEQ ID NO 4
 <211> LENGTH: 969
 <212> TYPE: DNA
 <213> ORGANISM: Clostridium perfringens
 <400> SEQUENCE: 4

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atgaaaagat taaaaattat ttcaattaca ctagtcttta caagtgtaat tagtacaagc    60
cttttttcaa ctcaaaactca agtttttgca agtgaattaa atgacataaa caaaattgag    120
ttgaaaaatc taagtggaga aataataaaa gaaaatggaa aggaagctat taaatatact    180
tctagtgata ccgcttcaca taaaggttgg aaggcaactt taagtggaac atttattgaa    240
gatcctcatt ctgataagaa aactgcttta ttaaatttag aaggatttat accttctgat    300
aaacagattt ttggttctaa atattacgga aaaatgaaat ggctgaaac ttatagaatt    360
aatgtaaaaa gtgctgatgt aaataataat ataaaaatag caaattctat tcctaaaaat    420
actatagata aaaaagatgt atctaattca attggttatt ctataggcgg taatatatct    480
gttgaaaggaa aaactgctgg tgctggaata aatgcttcat ataatgtcca aaatactata    540
agctatgaac aacctgattt tagaacaatt caaagaaaag atgatgcaaa cttagcatca    600
tgggatataa aatttgttga gactaaggac ggttataata tagattctta tcatgctatt    660
tatggaaatc aattattcat gaaatcaaga ttgtataata atggtgataa aaatttcaca    720
gatgatagag atttatcaac attaatttct ggtggatttt cacccaatat ggctttagca    780
ttaacagcac ctaaaaatgc taaagaatct gtaataatag ttgaatatca aagatttgat    840
aatgactata ttttaaattg ggaactact caatggcgag gaacaaacaa actttcgtca    900
acaagtgaat ataacgaatt tatgtttaa ataaattggc aagatcataa aatagaatat    960
tatctgtaa                                     969

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<210> SEQ ID NO 5
<211> LENGTH: 292
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutated sequence

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<400> SEQUENCE: 5

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Ser Glu Leu Asn Asp Ile Asn Lys Ile Glu Leu Lys Asn Leu Ser Gly
1             5             10            15

Glu Ile Ile Lys Glu Asn Gly Lys Glu Ala Ile Lys Tyr Thr Ser Ser
20            25            30

Asp Thr Ala Ser His Lys Gly Trp Lys Ala Thr Leu Ser Gly Thr Phe
35            40            45

Ile Glu Asp Pro His Ser Asp Lys Lys Thr Ala Leu Leu Asn Leu Glu
50            55            60

Gly Phe Ile Pro Ser Asp Lys Gln Ile Phe Gly Ser Lys Tyr Tyr Gly
65            70            75            80

Lys Met Lys Trp Pro Glu Thr Tyr Arg Ile Asn Val Lys Ser Ala Asp
85            90            95

Val Asn Asn Asn Ile Lys Ile Ala Asn Ser Ile Pro Lys Asn Thr Ile
100           105           110

Asp Lys Lys Asp Val Ser Asn Ser Ile Gly Tyr Ser Ile Gly Gly Asn
115           120           125

Ile Ser Val Glu Gly Lys Thr Ala Gly Ala Gly Ile Asn Ala Ser Tyr
130           135           140

Asn Val Gln Asn Thr Ile Ser Ala Glu Gln Pro Asp Phe Arg Thr Ile
145           150           155           160

Gln Arg Lys Asp Asp Ala Asn Leu Ala Ser Trp Asp Ile Lys Phe Val
165           170           175

Glu Thr Lys Asp Gly Tyr Asn Ile Asp Ser Tyr His Ala Ile Tyr Gly

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180	185	190
Asn Gln Leu Phe Met Lys Ser Arg Leu Tyr Asn Asn Gly Asp Lys Asn		
195	200	205
Phe Thr Asp Asp Arg Asp Leu Ser Thr Leu Ile Ser Gly Gly Phe Ser		
210	215	220
Pro Asn Met Ala Leu Ala Leu Thr Ala Pro Lys Asn Ala Lys Glu Ser		
225	230	235
Val Ile Ile Val Glu Tyr Gln Arg Phe Asp Asn Asp Tyr Ile Leu Asn		
245	250	255
Trp Glu Thr Thr Gln Trp Arg Gly Thr Asn Lys Leu Ser Ser Thr Ser		
260	265	270
Glu Tyr Asn Glu Phe Met Phe Lys Ile Asn Trp Gln Asp His Lys Ile		
275	280	285
Glu Tyr Tyr Leu		
290		
<210> SEQ ID NO 6		
<211> LENGTH: 292		
<212> TYPE: PRT		
<213> ORGANISM: Artificial Sequence		
<220> FEATURE:		
<223> OTHER INFORMATION: Mutated sequence		
<400> SEQUENCE: 6		
Ser Glu Leu Asn Asp Ile Asn Lys Ile Glu Leu Lys Asn Leu Ser Gly		
1	5	10
Glu Ile Ile Lys Glu Asn Gly Lys Glu Ala Ile Lys Tyr Thr Ser Ser		
20	25	30
Asp Thr Ala Ser His Lys Gly Trp Lys Ala Thr Leu Ser Gly Thr Phe		
35	40	45
Ile Glu Asp Pro His Ser Asp Lys Lys Thr Ala Leu Leu Asn Leu Glu		
50	55	60
Gly Phe Ile Pro Ser Asp Lys Gln Ile Phe Gly Ser Lys Tyr Tyr Gly		
65	70	75
Lys Met Lys Trp Pro Glu Thr Tyr Arg Ile Asn Val Lys Ser Ala Asp		
85	90	95
Val Asn Asn Asn Ile Lys Ile Ala Asn Ser Ile Ala Lys Asn Thr Ile		
100	105	110
Asp Lys Lys Asp Val Ser Asn Ser Ile Gly Tyr Ser Ile Gly Gly Asn		
115	120	125
Ile Ser Val Glu Gly Lys Thr Ala Gly Ala Gly Ile Asn Ala Ser Tyr		
130	135	140
Asn Val Gln Asn Thr Ile Ser Tyr Glu Gln Pro Asp Phe Arg Thr Ile		
145	150	155
Gln Arg Lys Asp Asp Ala Asn Leu Ala Ser Trp Asp Ile Lys Phe Val		
165	170	175
Glu Thr Lys Asp Gly Tyr Asn Ile Asp Ser Tyr His Ala Ile Tyr Gly		
180	185	190
Asn Gln Leu Phe Met Lys Ser Arg Leu Tyr Asn Asn Gly Asp Lys Asn		
195	200	205
Phe Thr Asp Asp Arg Asp Leu Ser Thr Leu Ile Ser Gly Gly Phe Ser		
210	215	220
Pro Asn Met Ala Leu Ala Leu Thr Ala Pro Lys Asn Ala Lys Glu Ser		
225	230	235
Val Ile Ile Val Glu Tyr Gln Arg Phe Asp Asn Asp Tyr Ile Leu Asn		

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	245	250	255	
Trp Glu Thr Thr Gln Trp Arg Gly Thr Asn Lys Leu Ser Ser Thr Ser				
	260	265	270	
Glu Tyr Asn Glu Phe Met Phe Lys Ile Asn Trp Gln Asp His Lys Ile				
	275	280	285	
Glu Tyr Tyr Leu				
	290			
<210> SEQ ID NO 7				
<211> LENGTH: 33				
<212> TYPE: DNA				
<213> ORGANISM: Artificial Sequence				
<220> FEATURE:				
<223> OTHER INFORMATION: Primer sequence				
<400> SEQUENCE: 7				
cccgggctcg agagtgaatt aaatgacata aac				33
<210> SEQ ID NO 8				
<211> LENGTH: 36				
<212> TYPE: DNA				
<213> ORGANISM: Artificial Sequence				
<220> FEATURE:				
<223> OTHER INFORMATION: Primer sequence				
<400> SEQUENCE: 8				
cccgggaagc ttttacagat aatattctat tttatg				36
<210> SEQ ID NO 9				
<211> LENGTH: 41				
<212> TYPE: DNA				
<213> ORGANISM: Artificial Sequence				
<220> FEATURE:				
<223> OTHER INFORMATION: Primer sequence				
<400> SEQUENCE: 9				
ggaacattta ttgaagctcc tcattctgat aagaaaactg c				41
<210> SEQ ID NO 10				
<211> LENGTH: 41				
<212> TYPE: DNA				
<213> ORGANISM: Artificial Sequence				
<220> FEATURE:				
<223> OTHER INFORMATION: Primer sequence				
<400> SEQUENCE: 10				
gcagttttct tatcagaatg aggagcttca ataaatgttc c				41
<210> SEQ ID NO 11				
<211> LENGTH: 44				
<212> TYPE: DNA				
<213> ORGANISM: Artificial Sequence				
<220> FEATURE:				
<223> OTHER INFORMATION: Primer sequence				
<400> SEQUENCE: 11				
gcaaattcta ttgctaaaaa tactatagat aaaaaagatg tatc				44
<210> SEQ ID NO 12				
<211> LENGTH: 44				
<212> TYPE: DNA				
<213> ORGANISM: Artificial Sequence				
<220> FEATURE:				
<223> OTHER INFORMATION: Primer sequence				

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<400> SEQUENCE: 12

gatacatctt ttttatctat agtattttta gcaatagaat ttgc 44

<210> SEQ ID NO 13

<211> LENGTH: 35

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 13

gatgtatcta attcaattgg tgcgtctata ggcgg 35

<210> SEQ ID NO 14

<211> LENGTH: 35

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 14

ccgcctatag acgcaccaat tgaattagat acatc 35

<210> SEQ ID NO 15

<211> LENGTH: 40

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 15

caattggtta ttctataggc gctaatatat ctgttgaagg 40

<210> SEQ ID NO 16

<211> LENGTH: 40

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 16

ccttcaacag atatattagc gcctatagaa taaccaattg 40

<210> SEQ ID NO 17

<211> LENGTH: 41

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 17

gtccaaaata ctataagcgc tgaacaacct gattttagaa c 41

<210> SEQ ID NO 18

<211> LENGTH: 41

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 18

gttctaaaat caggttggtc agcgcttata gtattttgga c 41

<210> SEQ ID NO 19

<211> LENGTH: 44

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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 19

ccaaaatact ataagctatg aagcacctga ttttagaaca attc          44

<210> SEQ ID NO 20
<211> LENGTH: 44
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 20

gaattgttct aaaatcaggt gcttcatagc ttatagtatt ttgg          44

<210> SEQ ID NO 21
<211> LENGTH: 40
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 21

ctataagcta tgaacaagct gattttagaa caattcaaag          40

<210> SEQ ID NO 22
<211> LENGTH: 40
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 22

ctttgaattg ttctaaaatc agcttggttca tagcttatag          40

<210> SEQ ID NO 23
<211> LENGTH: 37
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 23

caattattca tgaaatcagc attgtataat aatggtg          37

<210> SEQ ID NO 24
<211> LENGTH: 37
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer sequence

<400> SEQUENCE: 24

caccattatt atacaatgct gatttcatga ataattg          37

<210> SEQ ID NO 25
<211> LENGTH: 10
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (5)..(5)

```

-continued

<223> OTHER INFORMATION: Any amino acid other than Y

<400> SEQUENCE: 25

Asn Thr Ile Ser Xaa Glu Gln Pro Asp Phe
 1 5 10

<210> SEQ ID NO 26

<211> LENGTH: 20

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Mutant peptide fragment

<220> FEATURE:

<221> NAME/KEY: Xaa

<222> LOCATION: (10)..(10)

<223> OTHER INFORMATION: Any amino acid other than Y

<400> SEQUENCE: 26

Ser Tyr Asn Val Gln Asn Thr Ile Ser Xaa Glu Gln Pro Asp Phe Arg
 1 5 10 15

Thr Ile Gln Arg
 20

<210> SEQ ID NO 27

<211> LENGTH: 10

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Mutant peptide fragment

<220> FEATURE:

<221> NAME/KEY: Xaa

<222> LOCATION: (5)..(5)

<223> OTHER INFORMATION: Any amino acid other than P

<400> SEQUENCE: 27

Ala Asn Ser Ile Xaa Lys Asn Thr Ile Asp
 1 5 10

<210> SEQ ID NO 28

<211> LENGTH: 20

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Mutant peptide fragment

<220> FEATURE:

<221> NAME/KEY: Xaa

<222> LOCATION: (10)..(10)

<223> OTHER INFORMATION: any amino acid other than P

<220> FEATURE:

<221> NAME/KEY: Xaa

<222> LOCATION: (10)..(10)

<223> OTHER INFORMATION: Any amino acid other than P

<400> SEQUENCE: 28

Asn Asn Ile Lys Ile Ala Asn Ser Ile Xaa Lys Asn Thr Ile Asp Lys
 1 5 10 15

Lys Asp Val Ser
 20

<210> SEQ ID NO 29

<211> LENGTH: 68

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Polypeptide fragment

<400> SEQUENCE: 29

Asn Asn Asn Ile Lys Ile Ala Asn Ser Ile Pro Lys Asn Thr Ile Asp

-continued

```

1           5           10           15
Lys Lys Asp Val Ser Asn Ser Ile Gly Tyr Ser Ile Gly Gly Asn Ile
      20           25           30
Ser Val Glu Gly Lys Thr Ala Gly Ala Gly Ile Asn Ala Ser Tyr Asn
      35           40           45
Val Gln Asn Thr Ile Ser Tyr Glu Gln Pro Asp Phe Arg Thr Ile Gln
      50           55           60
Arg Lys Asp Asp
65

```

```

<210> SEQ ID NO 30
<211> LENGTH: 48
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Polypeptide fragment

```

```

<400> SEQUENCE: 30

```

```

Pro Lys Asn Thr Ile Asp Lys Lys Asp Val Ser Asn Ser Ile Gly Tyr
1           5           10           15
Ser Ile Gly Gly Asn Ile Ser Val Glu Gly Lys Thr Ala Gly Ala Gly
      20           25           30
Ile Asn Ala Ser Tyr Asn Val Gln Asn Thr Ile Ser Tyr Glu Gln Pro
      35           40           45

```

```

<210> SEQ ID NO 31
<211> LENGTH: 879
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant encoding DNA

```

```

<400> SEQUENCE: 31

```

```

agtgaattaa atgacataaa caaaattgag ttgaaaaatc taagtggaga aataataaaa      60
gaaaatggaa aggaagctat taaatatact tctagtata cgccttcaca taaagggttg      120
aaggcaactt taagtgaac atttattgaa gatcctcatt ctgataagaa aactgcttta      180
ttaaatntag aaggatttat accttctgat aaacagattt ttggttctaa atattacgga      240
aaaatgaaat ggctgaaac ttatagaatt aatgtaaaaa gtgctgatgt aaataataat      300
ataaaaaatag caaattctat tcttaaaaaa actatagata aaaaagatgt atctaattca      360
attggttatt ctataggcgg taatatatct gttgaaggaa aaactgctgg tgctggaata      420
aatgcttcat ataatgtcca aaatactata agcgctgaac aacctgattt tagaacaatt      480
caaagaaaag atgatgcaaa ttagcatca tgggatataa aatttggtga gactaaggac      540
ggttataata tagattctta tcatgtatt tatggaaatc aattattcat gaaatcaaga      600
ttgtataata atggtgataa aaatttcaca gatgatagag atttatcaac attaatttct      660
ggtggatttt caccatataat ggcttttagca ttaacagcac ctaaaaatgc taaagaatct      720
gtaataatag ttgaatatca aagatttgat aatgactata ttttaattg ggaaactact      780
caatggcgag gaacaaacaa acttttgtca acaagtgaat ataacgaatt tatgtttaaa      840
ataaattggc aagatcataa aatagaatat tatctgtaa                                879

```

```

<210> SEQ ID NO 32
<211> LENGTH: 879
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:

```

-continued

<223> OTHER INFORMATION: Mutant encoding DNA

<400> SEQUENCE: 32

agtgaattaa atgacataaa caaaattgag ttgaaaaatc taagtggaga aataataaaa	60
gaaaatggaa aggaagctat taaatatact tctagtata cgccttcaca taaaggttgg	120
aaggcaactt taagtggaac atttattgaa gatcctcatt ctgataagaa aactgcttta	180
ttaaatttag aaggatttat accttctgat aaacagattt ttggttctaa atattacgga	240
aaaatgaaat ggctgaaac ttatagaatt aatgtaaaa gtgctgatgt aaataataat	300
ataaaaaatag caaattctat tgctaaaaat actatagata aaaaagatgt atctaattca	360
attggttatt ctataggcgg taatatatct gttgaaggaa aaactgctgg tgctggaata	420
aatgcttcatt ataattgtca aaatactata agctatgaac aacctgattt tagaacaatt	480
caaagaaaag atgatgcaaa tttagcatca tgggatataa aatttggtga gactaaggac	540
ggttataata tagattctta tcatgctatt tatggaaatc aattattcat gaaatcaaga	600
ttgtataata atggtgataa aaatttcaca gatgatagag atttatcaac attaatctct	660
ggtggatttt caccatataat ggcttttagca ttaacagcac ctaaaaatgc taaagaatct	720
gtaataatag ttgaatatca aagatttgat aatgactata ttttaaattg ggaaactact	780
caatggcgag gaacaaacaa actttcgtca acaagtgaat ataacgaatt tatgtttaa	840
ataaattggc aagatcataa aatagaatat tatctgtaa	879

<210> SEQ ID NO 33

<211> LENGTH: 879

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Encoding mature NetB

<400> SEQUENCE: 33

agtgaattaa atgacataaa caaaattgag ttgaaaaatc taagtggaga aataataaaa	60
gaaaatggaa aggaagctat taaatatact tctagtata cgccttcaca taaaggttgg	120
aaggcaactt taagtggaac atttattgaa gatcctcatt ctgataagaa aactgcttta	180
ttaaatttag aaggatttat accttctgat aaacagattt ttggttctaa atattacgga	240
aaaatgaaat ggctgaaac ttatagaatt aatgtaaaa gtgctgatgt aaataataat	300
ataaaaaatag caaattctat tcctaaaaat actatagata aaaaagatgt atctaattca	360
attggttatt ctataggcgg taatatatct gttgaaggaa aaactgctgg tgctggaata	420
aatgcttcatt ataattgtca aaatactata agctatgaac aacctgattt tagaacaatt	480
caaagaaaag atgatgcaaa tttagcatca tgggatataa aatttggtga gactaaggac	540
ggttataata tagattctta tcatgctatt tatggaaatc aattattcat gaaatcaaga	600
ttgtataata atggtgataa aaatttcaca gatgatagag atttatcaac attaatctct	660
ggtggatttt caccatataat ggcttttagca ttaacagcac ctaaaaatgc taaagaatct	720
gtaataatag ttgaatatca aagatttgat aatgactata ttttaaattg ggaaactact	780
caatggcgag gaacaaacaa actttcgtca acaagtgaat ataacgaatt tatgtttaa	840
ataaattggc aagatcataa aatagaatat tatctgtaa	879

<210> SEQ ID NO 34

<211> LENGTH: 86

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

-continued

<220> FEATURE:

<223> OTHER INFORMATION: Polypeptide fragment

<400> SEQUENCE: 34

```

Tyr Asn Ile Asp Ser Tyr His Ala Ile Tyr Gly Asn Gln Leu Phe Met
1           5           10           15
Lys Ser Arg Leu Tyr Asn Asn Gly Asp Lys Asn Phe Thr Asp Asp Arg
          20           25           30
Asp Leu Ser Thr Leu Ile Ser Gly Gly Phe Ser Pro Asn Met Ala Leu
          35           40           45
Ala Leu Thr Ala Pro Lys Asn Ala Lys Glu Ser Val Ile Ile Val Glu
          50           55           60
Tyr Gln Arg Phe Asp Asn Asp Tyr Ile Leu Asn Trp Glu Thr Thr Gln
65           70           75           80
Trp Arg Gly Thr Asn Lys
          85

```

<210> SEQ ID NO 35

<211> LENGTH: 76

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Polypeptide fragment

<400> SEQUENCE: 35

```

Tyr His Ala Ile Tyr Gly Asn Gln Leu Phe Met Lys Ser Arg Leu Tyr
1           5           10           15
Asn Asn Gly Asp Lys Asn Phe Thr Asp Asp Arg Asp Leu Ser Thr Leu
          20           25           30
Ile Ser Gly Gly Phe Ser Pro Asn Met Ala Leu Ala Leu Thr Ala Pro
          35           40           45
Lys Asn Ala Lys Glu Ser Val Ile Ile Val Glu Tyr Gln Arg Phe Asp
          50           55           60
Asn Asp Tyr Ile Leu Asn Trp Glu Thr Thr Gln Trp
65           70           75

```

<210> SEQ ID NO 36

<211> LENGTH: 292

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Mutated polypeptide sequence

<400> SEQUENCE: 36

```

Ser Glu Leu Asn Asp Ile Asn Lys Ile Glu Leu Lys Asn Leu Ser Gly
1           5           10           15
Glu Ile Ile Lys Glu Asn Gly Lys Glu Ala Ile Lys Tyr Thr Ser Ser
          20           25           30
Asp Thr Ala Ser His Lys Gly Trp Lys Ala Thr Leu Ser Gly Thr Phe
          35           40           45
Ile Glu Asp Pro His Ser Asp Lys Lys Thr Ala Leu Leu Asn Leu Glu
          50           55           60
Gly Phe Ile Pro Ser Asp Lys Gln Ile Phe Gly Ser Lys Tyr Tyr Gly
65           70           75           80
Lys Met Lys Trp Pro Glu Thr Tyr Arg Ile Asn Val Lys Ser Ala Asp
          85           90           95
Val Asn Asn Asn Ile Lys Ile Ala Asn Ser Ile Pro Lys Asn Thr Ile
          100          105          110

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Asp Lys Lys Asp Val Ser Asn Ser Ile Gly Tyr Ser Ile Gly Gly Asn
   115                               120                               125

Ile Ser Val Glu Gly Lys Thr Ala Gly Ala Gly Ile Asn Ala Ser Tyr
   130                               135                               140

Asn Val Gln Asn Thr Ile Ser Tyr Glu Gln Pro Asp Phe Arg Thr Ile
   145                               150                               155                               160

Gln Arg Lys Asp Asp Ala Asn Leu Ala Ser Trp Asp Ile Lys Phe Val
   165                               170                               175

Glu Thr Lys Asp Gly Tyr Asn Ile Asp Ser Tyr His Ala Ile Ala Gly
   180                               185                               190

Asn Gln Leu Phe Met Lys Ser Arg Leu Tyr Asn Asn Gly Asp Lys Asn
   195                               200                               205

Phe Thr Asp Asp Arg Asp Leu Ser Thr Leu Ile Ser Gly Gly Phe Ser
   210                               215                               220

Pro Asn Met Ala Leu Ala Leu Thr Ala Pro Lys Asn Ala Lys Glu Ser
   225                               230                               235                               240

Val Ile Ile Val Glu Tyr Gln Arg Phe Asp Asn Asp Tyr Ile Leu Asn
   245                               250                               255

Trp Glu Thr Thr Gln Trp Arg Gly Thr Asn Lys Leu Ser Ser Thr Ser
   260                               265                               270

Glu Tyr Asn Glu Phe Met Phe Lys Ile Asn Trp Gln Asp His Lys Ile
   275                               280                               285

Glu Tyr Tyr Leu
   290

```

<210> SEQ ID NO 37

<211> LENGTH: 292

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Mutated polypeptide sequence

<400> SEQUENCE: 37

```

Ser Glu Leu Asn Asp Ile Asn Lys Ile Glu Leu Lys Asn Leu Ser Gly
  1                               5                               10                               15

Glu Ile Ile Lys Glu Asn Gly Lys Glu Ala Ile Lys Tyr Thr Ser Ser
   20                               25                               30

Asp Thr Ala Ser His Lys Gly Trp Lys Ala Thr Leu Ser Gly Thr Phe
   35                               40                               45

Ile Glu Asp Pro His Ser Asp Lys Lys Thr Ala Leu Leu Asn Leu Glu
   50                               55                               60

Gly Phe Ile Pro Ser Asp Lys Gln Ile Phe Gly Ser Lys Tyr Tyr Gly
   65                               70                               75                               80

Lys Met Lys Trp Pro Glu Thr Tyr Arg Ile Asn Val Lys Ser Ala Asp
   85                               90                               95

Val Asn Asn Asn Ile Lys Ile Ala Asn Ser Ile Pro Lys Asn Thr Ile
   100                              105                              110

Asp Lys Lys Asp Val Ser Asn Ser Ile Gly Tyr Ser Ile Gly Gly Asn
   115                               120                               125

Ile Ser Val Glu Gly Lys Thr Ala Gly Ala Gly Ile Asn Ala Ser Tyr
   130                               135                               140

Asn Val Gln Asn Thr Ile Ser Tyr Glu Gln Pro Asp Phe Arg Thr Ile
   145                               150                               155                               160

Gln Arg Lys Asp Asp Ala Asn Leu Ala Ser Trp Asp Ile Lys Phe Val
   165                               170                               175

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-continued

Glu Thr Lys Asp Gly Tyr Asn Ile Asp Ser Tyr His Ala Ile Tyr Gly
 180 185 190
 Asn Gln Leu Phe Met Lys Ser Ala Leu Tyr Asn Asn Gly Asp Lys Asn
 195 200 205
 Phe Thr Asp Asp Arg Asp Leu Ser Thr Leu Ile Ser Gly Gly Phe Ser
 210 215 220
 Pro Asn Met Ala Leu Ala Leu Thr Ala Pro Lys Asn Ala Lys Glu Ser
 225 230 235 240
 Val Ile Ile Val Glu Tyr Gln Arg Phe Asp Asn Asp Tyr Ile Leu Asn
 245 250 255
 Trp Glu Thr Thr Gln Trp Arg Gly Thr Asn Lys Leu Ser Ser Thr Ser
 260 265 270
 Glu Tyr Asn Glu Phe Met Phe Lys Ile Asn Trp Gln Asp His Lys Ile
 275 280 285
 Glu Tyr Tyr Leu
 290

<210> SEQ ID NO 38
 <211> LENGTH: 292
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Mutated polypeptide sequence

<400> SEQUENCE: 38

Ser Glu Leu Asn Asp Ile Asn Lys Ile Glu Leu Lys Asn Leu Ser Gly
 1 5 10 15
 Glu Ile Ile Lys Glu Asn Gly Lys Glu Ala Ile Lys Tyr Thr Ser Ser
 20 25 30
 Asp Thr Ala Ser His Lys Gly Trp Lys Ala Thr Leu Ser Gly Thr Phe
 35 40 45
 Ile Glu Asp Pro His Ser Asp Lys Lys Thr Ala Leu Leu Asn Leu Glu
 50 55 60
 Gly Phe Ile Pro Ser Asp Lys Gln Ile Phe Gly Ser Lys Tyr Tyr Gly
 65 70 75 80
 Lys Met Lys Trp Pro Glu Thr Tyr Arg Ile Asn Val Lys Ser Ala Asp
 85 90 95
 Val Asn Asn Asn Ile Lys Ile Ala Asn Ser Ile Pro Lys Asn Thr Ile
 100 105 110
 Asp Lys Lys Asp Val Ser Asn Ser Ile Gly Tyr Ser Ile Gly Gly Asn
 115 120 125
 Ile Ser Val Glu Gly Lys Thr Ala Gly Ala Gly Ile Asn Ala Ser Tyr
 130 135 140
 Asn Val Gln Asn Thr Ile Ser Tyr Glu Gln Pro Asp Phe Arg Thr Ile
 145 150 155 160
 Gln Arg Lys Asp Asp Ala Asn Leu Ala Ser Trp Asp Ile Lys Phe Val
 165 170 175
 Glu Thr Lys Asp Gly Tyr Asn Ile Asp Ser Tyr His Ala Ile Tyr Gly
 180 185 190
 Asn Gln Leu Phe Met Lys Ser Arg Leu Tyr Asn Asn Gly Asp Lys Asn
 195 200 205
 Phe Thr Asp Asp Arg Asp Leu Ser Thr Leu Ile Ser Gly Gly Phe Ser
 210 215 220
 Pro Asn Met Ala Leu Ala Leu Thr Ala Pro Lys Asn Ala Lys Glu Ser
 225 230 235 240

[illegible]

```
<210> SEQ ID NO 39
<211> LENGTH: 292
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutated polypeptide sequence
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<400> SEQUENCE: 39

[illegible]

-continued

<210> SEQ ID NO 40
<211> LENGTH: 10
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: X
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than Y
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than Y

<400> SEQUENCE: 40

Tyr His Ala Ile Xaa Gly Asn Gln Leu Phe
1 5 10

<210> SEQ ID NO 41
<211> LENGTH: 20
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: X
<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: Any amino acid other than Y
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: Any amino acid other than Y

<400> SEQUENCE: 41

Tyr Asn Ile Asp Ser Tyr His Ala Ile Xaa Gly Asn Gln Leu Phe Met
1 5 10 15

Lys Ser Arg Leu
20

<210> SEQ ID NO 42
<211> LENGTH: 10
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: X
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than R
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than R

<400> SEQUENCE: 42

Phe Met Lys Ser Xaa Leu Tyr Asn Asn Gly
1 5 10

<210> SEQ ID NO 43
<211> LENGTH: 20
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: X
<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: Any amino acid other than R
<220> FEATURE:
<221> NAME/KEY: Xaa

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<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: Any amino acid other than R

<400> SEQUENCE: 43

Tyr Gly Asn Gln Leu Phe Met Lys Ser Xaa Leu Tyr Asn Asn Gly Asp
1           5           10           15

Lys Asn Phe Thr
                20

```

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<210> SEQ ID NO 44
<211> LENGTH: 10
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: X
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than W
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than W

<400> SEQUENCE: 44

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Tyr Ile Leu Asn Xaa Glu Thr Thr Gln Trp
1           5           10

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<210> SEQ ID NO 45
<211> LENGTH: 20
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: X
<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: Any amino acid other than W.
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: Any amino acid other than W.

<400> SEQUENCE: 45

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Arg Phe Asp Asn Asp Tyr Ile Leu Asn Xaa Glu Thr Thr Gln Trp Arg
1           5           10           15

Gly Thr Asn Lys
                20

```

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<210> SEQ ID NO 46
<211> LENGTH: 10
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant peptide fragment
<220> FEATURE:
<221> NAME/KEY: X
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than W
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than W
<220> FEATURE:
<221> NAME/KEY: Xaa
<222> LOCATION: (5)..(5)
<223> OTHER INFORMATION: Any amino acid other than W

<400> SEQUENCE: 46

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Glu Thr Thr Gln Xaa Arg Gly Thr Asn Lys

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-continued

1 5 10

<210> SEQ ID NO 47
 <211> LENGTH: 20
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Mutant peptide fragment
 <220> FEATURE:
 <221> NAME/KEY: X
 <222> LOCATION: (10)..(10)
 <223> OTHER INFORMATION: Any amino acid other than W
 <220> FEATURE:
 <221> NAME/KEY: Xaa
 <222> LOCATION: (10)..(10)
 <223> OTHER INFORMATION: Any amino acid other than W

<400> SEQUENCE: 47

Tyr	Ile	Leu	Asn	Trp	Glu	Thr	Thr	Gln	Xaa	Arg	Gly	Thr	Asn	Lys	Leu
1				5				10					15		
Ser	Ser	Thr	Ser												
				20											

<210> SEQ ID NO 48
 <211> LENGTH: 879
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Mutant encoding DNA

<400> SEQUENCE: 48

agtgaattaa atgacataaa caaaattgag ttgaaaaatc taagtggaga aataataaaa	60
gaaaatggaa aggaagctat taaatatact tctagtata cgccttcaca taaaggttgg	120
aaggcaactt taagtggaac atttattgaa gatcctcatt ctgataagaa aactgcttta	180
ttaaatttag aaggatttat accttctgat aaacagattt ttggttctaa atattacgga	240
aaaatgaaat ggctgaaac ttatagaatt aatgtaaaa gtgctgatgt aaataataat	300
ataaaaaatag caaattctat tcttaaaaat actatagata aaaaagatgt atctaattca	360
attggttatt ctataggcgg taatatatct gttgaaggaa aaactgctgg tgctggaata	420
aatgcttcac ataattgcca aaatactata agctatgaac aacctgattt tagaacaatt	480
caaagaaaag atgatgcaaa ttagcatca tgggatataa aatttgttga gactaaggac	540
ggttataata tagattctta tcatgctatt gctggaaatc aattattcat gaaatcaaga	600
ttgtataata atggtgataa aaatttcaca gatgatagag atttatcaac attaatctct	660
ggtggatttt caccatataat ggcttttagca ttaacagcac ctaaaaatgc taaagaatct	720
gtaataatag ttgaatatca aagatttgat aatgactata ttttaaattg ggaaactact	780
caatggcgag gaacaaacaa actttcgtca acaagtgaat ataacgaatt tatgtttaaa	840
ataaattggc aagatcataa aatagaatat tatctgtaa	879

<210> SEQ ID NO 49
 <211> LENGTH: 879
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Mutant encoding DNA

<400> SEQUENCE: 49

agtgaattaa atgacataaa caaaattgag ttgaaaaatc taagtggaga aataataaaa	60
gaaaatggaa aggaagctat taaatatact tctagtata cgccttcaca taaaggttgg	120

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aaggcaactt taagtgaac atttattgaa gatcctcatt ctgataagaa aactgcttta 180
ttaaatntag aaggatttat accttctgat aaacagattt ttggttctaa atattacgga 240
aaaatgaaat ggctgaaac ttatagaatt aatgtaaaaa gtgctgatgt aaataataat 300
ataaaaaatag caaattctat tcctaaaaat actatagata aaaaagatgt atctaattca 360
attggttatt ctataggcgg taatatatct gttgaaggaa aaactgctgg tgctggaata 420
aatgcttcat ataatgtcca aaatactata agctatgaac aacctgattt tagaacaatt 480
caaagaaaag atgatgcaaa ttagcatca tgggatataa aatttgttga gactaaggac 540
ggttataata tagattctta tcatgctatt tatggaaatc aattattcat gaaatcagca 600
ttgtataata atggtgataa aaatttcaca gatgatagag atttatcaac attaatctct 660
ggtggatttt cacccaatat ggctttagca ttaacagcac ctaaaaatgc taaagaatct 720
gtaataatag ttgaatatca aagatttgat aatgactata ttttaaattg ggaaactact 780
caatggcgag gaacaaacaa actttcgtca acaagtgaat ataacgaatt tatgtttaa 840
ataaattggc aagatcataa aatagaatat tatctgtaa 879

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<210> SEQ ID NO 50
<211> LENGTH: 879
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant encoding DNA

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<400> SEQUENCE: 50

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agtgaattaa atgacataaa caaaattgag ttgaaaaatc taagtggaga aataataaaa 60
gaaaatggaa aggaagctat taaatatact tctagtata ccgcttcaca taaaggttgg 120
aaggcaactt taagtgaac atttattgaa gatcctcatt ctgataagaa aactgcttta 180
ttaaatntag aaggatttat accttctgat aaacagattt ttggttctaa atattacgga 240
aaaatgaaat ggctgaaac ttatagaatt aatgtaaaaa gtgctgatgt aaataataat 300
ataaaaaatag caaattctat tcctaaaaat actatagata aaaaagatgt atctaattca 360
attggttatt ctataggcgg taatatatct gttgaaggaa aaactgctgg tgctggaata 420
aatgcttcat ataatgtcca aaatactata agctatgaac aacctgattt tagaacaatt 480
caaagaaaag atgatgcaaa ttagcatca tgggatataa aatttgttga gactaaggac 540
ggttataata tagattctta tcatgctatt tatggaaatc aattattcat gaaatcaaga 600
ttgtataata atggtgataa aaatttcaca gatgatagag atttatcaac attaatctct 660
ggtggatttt cacccaatat ggctttagca ttaacagcac ctaaaaatgc taaagaatct 720
gtaataatag ttgaatatca aagatttgat aatgactata ttttaaattg ggaaactact 780
caatggcgag gaacaaacaa actttcgtca acaagtgaat ataacgaatt tatgtttaa 840
ataaattggc aagatcataa aatagaatat tatctgtaa 879

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<210> SEQ ID NO 51
<211> LENGTH: 879
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Mutant encoding DNA

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<400> SEQUENCE: 51

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agtgaattaa atgacataaa caaaattgag ttgaaaaatc taagtggaga aataataaaa 60

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-continued

gaaaatggaa aggaagctat taaatatact tctagtata cgccttcaca taaagggttg	120
aaggcaactt taagtgaac atttattgaa gatcctcatt ctgataagaa aactgcttta	180
ttaaatttag aaggatttat accttctgat aaacagattt ttggttctaa atattacgga	240
aaaatgaaat ggctgaaac ttatagaatt aatgtaaaaa gtgctgatgt aaataataat	300
ataaaaaatag caaattctat tcctaaaaat actatagata aaaaagatgt atctaattca	360
attggttatt ctataggcgg taatatatct gttgaaggaa aaactgctgg tgctggaata	420
aatgcttcat ataatgtcca aaatactata agctatgaac aacctgattt tagaacaatt	480
caaagaaaag atgatgcaaa tttagcatca tgggatataa aatttgttga gactaaggac	540
ggttataata tagattctta tcctgctatt tatggaaatc aattattcat gaaatcaaga	600
ttgtataata atggtgataa aaatttcaca gatgatagag atttatcaac attaatttct	660
ggtggatttt caccacaat atgcttttagca ttaacagcac ctaaaaatgc taaagaatct	720
gtaataatag ttgaatatca aagatttgat aatgactata ttttaaattg ggaaactact	780
caagcgcgag gaacaaacaa actttcgtca acaagtgaat ataacgaatt tatgtttaa	840
ataaattggc aagatcataa aatagaatat tatctgtaa	879

The invention claimed is:

1. A polypeptide having at least 60% sequence identity to SEQ ID NO: 1 and comprising the amino acid sequence ETTQXRGTNK (SEQ ID NO: 46) where "X" is any amino acid other than W, the polypeptide having reduced toxicity compared with the toxicity of SEQ ID NO: 1.

2. The polypeptide according to claim 1 comprising the amino acid sequence ETTQXRGTNK (SEQ ID NO: 46) or YILNWETTQXRGTNKLST (SEQ ID NO: 47), where "X" is any amino acid other than W.

3. The polypeptide according to claim 1 wherein "X" is amino acid alanine.

4. The polypeptide according to claim 1 having the amino acid sequence SEQ ID NO: 39.

5. A cell comprising a polypeptide according to claim 1.

6. A composition comprising a polypeptide according to claim 1.

7. A method of reducing the risk of an infection by *Clostridium perfringens*, by inducing an immune response through administering to the subject a protective amount of a polypeptide according to claim 1.

8. A kit comprising a polypeptide according claim 1.

* * * * *